Effect of methanolic extract of Asparagus racemosus Willd. on lipopolysaccharide induced-oxidative stress in rats

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Abstract: Lipopolysaccharide (LPS) induced oxidative stress and impairment of normal physiological function generally categorized by increased anxiety and reduced mobility. Therefore, the present study was to find out the effect Methanolic extract of Asparagus racemosus (MEAR) in lipopolysaccharide (LPS)-induced oxidative stress in rats. LPS-induced oxidative stress in rats was measured by locomotor activity by photoactometer test, anxiety with elevated plus maze test and also studied the oxidative stress markers, nitric oxide and cytokines. The obtained data shows that LPS markedly exhausted (p<0.001) brain-reduced glutathione (GSH), superoxide dismutase (SOD) and catalase (CAT) significantly increased (p<0.001) the level of malondialdehyde (MDA), nitric oxide and the activity of cytokines in the brain. MEAR supplementation resulted in normalization of brain GSH and CAT and SOD and decreases in the levels of MDA with reduction of nitric oxide and cytokines in the brain. The action of the extract at dose of 200 mg/kg was almost similar to the standard drug, quercetin (100mg/kg, p.o.). These present study conclude that MEAR administration significantly (P<0.05) reduced LPS- induced oxidative-stress and intensely suggest that Asparagus racemosus Willd. is a functionally newer type of cerebroprotective agent.

Keywords: Oxidative stress, Asparagus racemosus, lipopolysaccharide, cytokine, endotoxin.

INTRODUCTION

Lipopolysaccharide (LPS) is a gram-negative bacterial endotoxin and a key element that gives to organ failure, including brain injury (Kheir-Eldin et al., 2001). The degree of oxidative stress and the severity of resulting organ damage might depend on the imbalance between excess production of reactive oxygen species (ROS) and antioxidant defensive mechanism (Tanguy et al., 2003). LPS binding to immune cells initiates a cascade of happenings that up-regulate manifestation of the inflammatory cytokines including tissue necrosis factor -α (TNF-α), interleukin-6, (IL-6) and interleukin-1β (IL-1β). TNF-α and other cytokines stimulates the production of ROS and reactive nitrogen intermediates (RNIs) by activated macrophages causing brain due to the oxidative stress (Victor and Fuente, 2003). Additionally, LPS induces passage of stimulated polymorph nuclear leukocytes (PMNs) into the brain, which creates another source of free radicals. The oxidative stress thus generated, induces a rapid alteration in the antioxidant generation systems by depleting the cellular stores of endogenous antioxidants such as superoxide dismutase (SOD), catalase, glutathione (Richard et al., 1990; Wojewoda et al., 2010; Sangeeta et al., 2011). Asparagus racemosus Willd. is a main therapeutic plant of tropical and subtropical region of India and identified as rasayana and commonly called shatavari. In India its widely used in gastrointestinal disorder, as antitussive, as an immunostimulant and as antispasmodic (Bopana and Saxena, 2007). Asparagus racemosus has been used in Ayurveda as a galactagogue, anodyne, antispasmodic, diuretic, antiurolithiatic activity, hepatoprotective action and in diabetic nephropathy (Sharma et al., 2000; Jagannath et al., 2012; Acharya et al., 2012; Somania et al., 2012). The present study was aimed to evaluate the activity of methanolic extract of Asparagus racemosus on lipopolysaccharide induced oxidative stress in rats.

MATERIALS AND METHODS

Plants materials
Roots of Asparagus racemosus Willd. was collected from Hamdard Dawakhana, Lucknow, India. The plant material was identified and authenticated by a taxonomist of National Botanical Research Institute (NBRI), Lucknow, India and the voucher specimen number NBRI-SOP-202 was deposited in the departmental herbarium.

Chemicals
All chemicals used were of analytical grade. Thiobarbituric acid (TBA); 2,6-di-tert-butyl-4-hydroxytoluene (BHT); trichloroacetic acid (TCA); Hydrogen peroxide (H2O2); EDTA; Tris buffer; Potassium dihydrogen ortho phosphate; Disodium hydrogen ortho phosphate were obtained from CDH, Mumbai, NADPH; DTNB were obtained from Hi Media, Mumbai, cytokine ELISA kits obtained from eBioscience and Cayman Chemical USA, Lipopolysaccharide Serotype E. coli

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0111: B4 were obtained from Sigma Chemicals, USA and standard drug quercetin were obtained from Total herb solution Mumbai.

Animals
Adult male (8 weeks) Sprague Dawley (SD) rats weighing 150-160g were used in this study. The animals were procured from the Division of Laboratory Animal, Central Drug Research Institute (CDRI), Lucknow, India. They were kept in polycrylic cages in-group of seven and maintained under standard housing condition (room temperature 25±1°C and humidity 60-65%) with 12h light and dark cycle. The food and water were available ad libitum. Experiments were performed as per internationally followed ethical standards. This study was approved by Institutional Animal Ethics Committee (IAEC) of Integral University and CPCSEA No- IU/Pharm/Ph.D/CPCSEA/12/03.

Preparation of plant extract
The root of Asparagus racemosus was air-dried and dried materials were pulverized in a laboratory mill to a moderately fine powder. The powder was filled into soxhlet apparatus and exposed to hot continuous percolation using methanol (95% v/v) as solvent. The extract was concentrated under vacuum and dried in a vacuum desiccator (yield 10.4% w/w).

Acute toxicity studies
Acute toxicity: The acute toxicity of the extract of Root of Asparagus racemosus Willd. was evaluated in mice using the up and down procedure (OECD, 2001). Mice received methanolic extract at various doses (500-2000mg/kg) orally by gavage. The animals were observed for abnormal behaviour continuously for the first 4h after dosing. Finally, the number of survivors was noted after 24 h.

Experimental design
Animals were randomly divided into seven groups as follows: Group I (Control group) 1% Carboxy methyl cellulose (CMC) 5ml/kg p.o. once a day, and one dose of normal saline at day 21 from the start of the experiment (4 mg/kg i.p.). Group II (LPS group) 1% CMC 5ml/kg, p.o. once a day, and one dose of LPS Serotype E. coli 0111: B4 at day 21 from the start of the experiment (4 mg/kg, i.p.). Groups III (standard group) Quercetin 100 mg/kg, p.o. daily, and one dose of LPS at day 21 from the start of the experiment (4mg/kg, i.p.). Groups IV and V (Drugs treated groups) 100mg/kg and 200mg/kg, p.o. daily, and one dose of LPS at day 21 from the start of the experiment (4mg/kg i.p.) respectively. Groups VI 200 mg/kg of test drug and 100mg/kg standard drug p.o. and one dose of LPS at day 21 from the start of the experiment (4mg/kg, i.p.). VII (perse group) 200mg/kg, p.o. test drug daily for 21 days.

Anxiety was assessed in the elevated plus mazes 3 h after the LPS or saline administration on 21 day. Motor activity was assessed in the photoactometer test 4h after the LPS or saline administration on 21 day. Rats were transcardially perfused with cold saline followed by 4 % formalin in phosphate buffer-saline (o.1 M; pH 7.4). The brains were removed and stored -80°C and later use for biochemical estimation.

Parameters investigated
(i) Behavioural tests
Elevated plus maze test (Corbett et al., 1986). and Photoactometer test (Ahmad et al., 2012) carried out by reported procedure.

(ii) Biochemical estimations
Estimation of Thio Barbituric Acid Reactive Substances (TBARS), (Ohkawa et al., 1979) glutathione(GSH) (Sedlak and Lindsay,1968), Catalase (CAT) (Clairbone, 1985), Super oxide dismutase (SOD) (Marklund and Marklund, 1974) and NO and cytokine (eBioscience and Cayman Chemical USA) carried out according to the manufacturer's instructions.

STATISTICAL ANALYSIS
The values were expressed as mean ±SEM. Statistical analysis was done by one way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparison test. P values<0.05 were considered as significant.

RESULTS
Assessment of elevated maze test
LPS induced an anxiogenic effect in only LPS treated group and significantly (P<0.05) reduced the time spent in open arm in plus maze. Oral administration of MEAR for 21 days increased time spent in open arm and significantly reversed the effect of LPS (table 1).

Assessment of photoactometer response
LPS suppress Locomotor activity was increased by Asparagus racemosus root extract and significantly (P<0.05) increased (table 1).

Reduced glutathione (GSH) content in brain
In the present study, LPS administration (4mg/kg, i.p, once) resulted in significant depletion of brain GSH compared to the corresponding control value. Administration of 100mg/ kg and 200 mg/kg MEAR as an oral daily for 21 days as well as LPS (4mg/kg, i.p, once at the 21day) markedly increased brain (p<0.001) GSH content as it reached normal levels (table 2).

Malondialdehyde (MDA) content in brain
By evaluation of MDA (as indicator of lipid peroxides formation) in rat brain, the obtained data (table 2) explain that LPS administration (4mg/kg, i.p, once) into rats...
markedly increased the level of brain MDA compared to the corresponding control animals. However animals administered both MEAR (100mg/kg, p.o daily) in 21 successive days and injected with a single dose of LPS (4mg/kg, i.p) at the 21st day showed marked reduction (p<0.001) in brain MDA content in comparison with only LPS-administered animals.

**Superoxide dismutase (SOD) content in brain**
LPS administration decreases the level of SOD in to rats. Animal administered MEAR 100mg/kg and 200 mg/kg p.o. daily for 21th days and injected with single dose of LPS (4mg/kg i.p) at the 21st day marked increase (p<0.001) in CAT in comparison with only LPS-administered animals.

**Catalase (CAT) content in brain**
LPS administration decreases the level of CAT in to rats. Animal administered MEAR 100mg/kg and 200mg/kg p.o. daily for 21th days and injected with single dose of LPS (4mg/kg i.p) at the 21st day marked increase (p<0.001) in CAT in comparison with only LPS-administered animals.

**Nitric oxide content in brain**
In the rats who were treated with Asparagus racemosus 100mg/kg and 200mg/kg, the levels of NO were significantly reduced as compared to disease control (table 3) shows the effect of the significant and dose dependent recovery on the LPS induced elevation of the NO levels in animals.

**Cytokines content in brain**
In the rats who were pretreated with Asparagus racemosus, the levels of cytokines were significantly reduced (p<0.001) as compared to disease control (table 3) shows the effect of the significant and dose dependent recovery on the LPS induced elevation of the cytokines levels in animals.

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**Table 1: Assessment of Plus maze and Photoactometer**

<table>
<thead>
<tr>
<th>Group</th>
<th>Drugs treatment</th>
<th>Time duration in open arm in sec</th>
<th>No of entries in open arm</th>
<th>Locomotor activity</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Control</td>
<td>204.6±5.115#</td>
<td>6.6±0.509#</td>
<td>178±10.677##</td>
</tr>
<tr>
<td>II</td>
<td>LPS</td>
<td>199.6±3.558*</td>
<td>2.00±0.136#</td>
<td>121±6.403*</td>
</tr>
<tr>
<td>III</td>
<td>QT</td>
<td>180.8±3.338#</td>
<td>5.2±0.374#</td>
<td>170±8.509###</td>
</tr>
<tr>
<td>IV</td>
<td>AR1</td>
<td>140.4±3.265#</td>
<td>2.4±0.244 ns</td>
<td>123±5.380ns</td>
</tr>
<tr>
<td>V</td>
<td>AR2</td>
<td>150±3.536#</td>
<td>4.00±0.362##</td>
<td>169.8±8.777###</td>
</tr>
<tr>
<td>VI</td>
<td>AR2+QT</td>
<td>181.4±2.943#</td>
<td>5.6±0.244#</td>
<td>171±8.741###</td>
</tr>
<tr>
<td>VII</td>
<td>AR2+perse</td>
<td>198.8±3.513</td>
<td>7±0.316</td>
<td>176.4±10.815</td>
</tr>
</tbody>
</table>

**Table 2: Assessment of oxidative stress markers**

<table>
<thead>
<tr>
<th>Drugs treatment</th>
<th>GSH (µg/mg protein)</th>
<th>CAT (nmol H2O2/mg protein)</th>
<th>SOD (Units/mg protein)</th>
<th>TBARS (nmoles MDA/mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>3.452±0.080#</td>
<td>22.2±0.860#</td>
<td>2.604±0.106#</td>
<td>3.658±0.076#</td>
</tr>
<tr>
<td>LPS</td>
<td>1.51±0.050*</td>
<td>9.00±0.707*</td>
<td>1.46±0.1281*</td>
<td>7.56±0.096*</td>
</tr>
<tr>
<td>QT</td>
<td>3.13±0.070#</td>
<td>19.4±0.500#</td>
<td>2.42±0.128#</td>
<td>3.79±0.051#</td>
</tr>
<tr>
<td>AR1</td>
<td>1.894±0.110ns</td>
<td>11.4±0.678ns</td>
<td>1.76±0.114ns</td>
<td>7.06±0.092##</td>
</tr>
<tr>
<td>AR2</td>
<td>1.972±0.037##</td>
<td>13.2±0.66##</td>
<td>2.02±0.073##</td>
<td>5.26±0.136#</td>
</tr>
<tr>
<td>AR2+QT</td>
<td>3.35±0.065#</td>
<td>20.4±0.678#</td>
<td>2.46±0.0812#</td>
<td>3.704±0.011#</td>
</tr>
<tr>
<td>AR2+perse</td>
<td>3.516±0.036#</td>
<td>22.8±0.583</td>
<td>2.68±0.086</td>
<td>3.746±0.130</td>
</tr>
</tbody>
</table>

**Table 3: Assessment of cytokines level and nitric oxide in brain**

<table>
<thead>
<tr>
<th>Group</th>
<th>Drugs treatment</th>
<th>TNF-α Pg/ml</th>
<th>IL-6 Pg/ml</th>
<th>IL-1β Pg/ml</th>
<th>NO (µmol nitrite/mg of wet tissue)</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Control</td>
<td>33.00±0.707#</td>
<td>138.4±2.657#</td>
<td>38.4±1.364#</td>
<td>3.38±0.086#</td>
</tr>
<tr>
<td>II</td>
<td>LPS</td>
<td>52.8±1.71*</td>
<td>208±5.762*</td>
<td>79.4±1.503*</td>
<td>8.42±0.0712*</td>
</tr>
<tr>
<td>III</td>
<td>QT</td>
<td>37.00±1.09#</td>
<td>144±2.429#</td>
<td>43.4±1.288#</td>
<td>5.866±0.1388#</td>
</tr>
<tr>
<td>IV</td>
<td>AR1</td>
<td>42.8±1.715#</td>
<td>185.6±1.806#</td>
<td>69.8±1.772#</td>
<td>7.01±0.135#</td>
</tr>
<tr>
<td>V</td>
<td>AR2</td>
<td>40.8±1.68#</td>
<td>182.8±5.809#</td>
<td>65.4±1.240#</td>
<td>6.322±0.066#</td>
</tr>
<tr>
<td>VI</td>
<td>AR2+QT</td>
<td>35.4±0.927#</td>
<td>143.4±2.804#</td>
<td>43.6±1.631#</td>
<td>5.78±0.105#</td>
</tr>
<tr>
<td>VII</td>
<td>AR2+perse</td>
<td>34.4±1.50</td>
<td>138.6±2.315</td>
<td>38±0.707</td>
<td>3.57±0.129</td>
</tr>
</tbody>
</table>

LPS-Lipopolysacchride, AR1- Lower dose of Asparagus racemosus extract, AR2- Higher dose of Asparagus racemosus extract, QT-Quercetin Results are expressed as mean ± SEM. The results were analyzed by Analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparison test. n=6 (Number of animal per group) # = p<0.001, ##= p<.01, ### = p<0.05, * = p<0.001, # Vs. GroupII, *Vs. GroupI
DISCUSSION

Medicinal plant and their products are accepted for their influential pharmacological activities in health and disease treatment. They are able to exert antioxidant, antiradical and hepatoprotective activities (Amor et al., 2009). In the present study, administration of LPS to rats resulted in development of oxidative stress which leads to suppressed locomotor and exploratory activity, induced an anxiogenic response and damage in brain tissue in rats. This effect was pointed out by an increase in the concentration of lipid peroxidation, cytokines, nitric oxide and decrease in the concentration of the GSH, SOD and Catalase. It is well accepted that LPS causes oxidative stress by intensification of proinflammatory cytokines production and by inducing the generation of ROS by different mechanisms (Victor et al., 2001; Zhao et al., 2008; Jaworek et al., 2007). Lipid peroxidation causes tissues injury by inactivation of membrane enzymes and receptors, depolymerization of polysaccharides as well as protein cross-linking and fragmentation (Luqman and Rizvi, 2006). This results in severe metabolic dysfunction and loss of cell integrity as well as genomic stability. Brain tissues have rich polyunsaturated fatty acids and are known for its high oxygen uptake. Therefore, it is more susceptible to oxidative stress than other tissues (Rozenberg et al., 2006). Both the doses of extract of Asparagus racemosus Willd. root (MEAR) significantly decreased brain cytokines levels after 6 h of LPS administration as compared to rats treated with disease control. In the present investigation increased the levels of GSH, SOD, catalase and decrease the level of lipid peroxidation cytokines and nitric oxide significantly in the LPS-challenged animals after the Root of Asparagus racemosus supplementation. Improvement of antioxidant status in root of Asparagus racemosus Willd. supplemented groups is in agreement with earlier findings (Gulec et al., 2006; Bansal et al., 2005). Increased TNF-α has been repeatedly shown to play a pivotal role in LPS-induced brain injury. TNF-α is a multifunctional cytokine secreted by activated macrophages, monocytes, neutrophils and Natural killer-cells. In accession to its direct cytotoxic effects, it is able to accelerate chemokines macrophage chemotactic protein-1 and vascular cell adhesion molecule-1, which is the key to hyper inflammation and ensuing brain damage. Asparagus racemosus Willd. is a medicinal plant with well-known antioxidant property (Hussain et al., 2011). Scientific evaluation of this claim using experimental model of LPS induced oxidative stress in rats was ascertained in this study. Asparagus racemosus Willd. contains steroidal saponins that are present in the root (Nandgopal et al., 2011). This constituent might be responsible for this effect. In conclusion, Oral administration of methanolic extract of Asparagus racemosus Willd. root (MEAR) protected rats from LPS induced brain injury. The protection may be due to the decrease of oxidative stress, which occurs by alteration in levels of antioxidant enzymes in oxidative stress rats. These observations propose that MEAR may be clinically viable protection against variety of conditions where cellular damage is a consequence of oxidative stress. In conclusion, the present study provides experimental evidence for methanolic extract of Asparagus racemosus Willd. as a cerebroprotective agent.

REFERENCES


