Protective effect of *Cnidoscolus aconitifolius* leaves against diclofenac-induced gastric mucosal damage

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**Abstract:** *Cnidoscolus aconitifolius* is one of the plants used in folk medicine for the treatment of peptic ulcer. However, the present study was designed to validate the gastro-protective effect of *Cnidoscolus aconitifolius* leaves using diclofenac-induced ulcer model. The gastro-protective effect of *Cnidoscolus aconitifolius* leaves was evaluated by employing diclofenac-induced gastric ulcer in rats. Gastric mucosal lesions were induced by oral administration of diclofenac in rats. The pH of gastric juice from pylorus-ligated rats was measured. Histological examinations of the gastric tissues were also carried out. The present findings showed that *Cnidoscolus aconitifolius* leaf methanol extract exerted gastro-protective activity, as demonstrated by its consistent significant and dose dependent increase in mean ulcer index (MUI) inhibition against gastric mucosa damage induced by diclofenac treatment. Histological investigation of the gastric lesions showed that the plant stimulates the scarring cicatrizing process and thereby maintaining the architecture of the mucosal surfaces. The gastro-protective effect observed could be related to the therapeutic properties of the phytoconstituents of *Cnidoscolus aconitifolius* extract. *Cnidoscolus aconitifolius* leaves possess gastro-protective property in a dose dependent manner.

**Keywords:** *Cnidoscolus aconitifolius*, phytochemicals, ulcer, diclofenac-induced ulcer model, ulcer index, histology.

**INTRODUCTION**

According to Rezq and Elmalh (2010), peptic ulcer disease is a problem of the gastrointestinal tract characterized by mucosal damage secondary to pepsin and gastric acid secretion. The event of gastric ulcer mostly involves an imbalance between defensive (protective) factors such as (Mucus production, bicarbonate, prostaglandins, nitric oxide and blood flow to mucosa) and aggressive (hostile) factors such as (gastric acid, pepsin, *Helicobacter pylori* and non-steroidal anti-inflammatory drugs (NSAIDs) in the mucosal membrane of stomach and duodenum (Bigheti et al., 2005). Ulcerative lesions of gastrointestinal tract is a major side effect associated with the use of NSAIDs, stress, alcohol and ischemic reperfusion and secretion of gastric acid is central component in the pathogenesis (Luiz-Ferrerra et al., 2010).

It has been reported that along with cyclooxygenase-dependent and independent mechanisms, various factors including stimulation of gastric acid production, inflammatory cells infiltration, cytokines, mucosal blood flow, and free radicals contribute to the development of NSAID-induced gastric mucosal damage. However, gastrointestinal toxicity associated with NSAIDs is an important medical problem and thus, the ability of NSAIDs to inhibit cyclooxygenase (COX) activities contributes to the cytotoxicity of these drugs in the gastrointestinal tract. Conventional anti-ulcer drugs such as histamine antagonists (cimetidine and ranitidine etc), proton pump inhibitors (omeprazol and benzimidazole etc), prostaglandin analogues and drugs affecting mucosal barrier are currently in use. Despite the therapeutic effects of these drugs, they are usually expensive and occasionally associated with relapse and adverse effects during ulcer therapy (Dharmani and Palit, 2006). This led to renewed interest in the search for new anti-ulcer drugs from natural sources.

Natural products, such as plants extract, either as pure compounds or as standardized extracts, provide limitless opportunities for new drug discoveries because of the unmatched availability of chemical diversity (Cos et al., 2006; Sasidharan et al., 2011). According to Moghadamtousi et al. (2015), natural products from medicinal plants have been the basis of many traditional medicine systems throughout the world. *Cnidoscolus aconitifolius* is a perennial shrub which belongs to the family Euphorbiaceae. It is commonly found in the tropic and sub-tropical regions worldwide, including Africa, South of Sahara, North and South America, India, etc. It is commonly eaten as vegetable in soup in south western Nigeria where it is called “Iyana Ipaja”, “hospital is too far” (Oyagbemi and Otedola, 2013). Hepatoprotective, anti-inflammatory, and analgesic properties of *Cnidoscolus aconitifolius* leaves have been reported as well as its effect against multi-drug resistant microorganism (Adaramoye et al., 2011). In addition, it has antidiabetic property and antibacterial activities (Azeez et al., 2010; Awoyinka et al., 2007). Its use in the treatment of obesity, acne, kidney stones and eye problems has also been reported (Kuti and Torres, 1996). Despite the ethnomedicinal importance of *Cnidoscolus aconitifolius*, no study has been carried out...
to validate the usefulness of *Cnidoscolus aconitifolius* in treatment of peptic ulcer.

**Table 1: Phytochemical Composition of the Methanol Extract of *Cnidoscolus aconitifolius* Leaves**

<table>
<thead>
<tr>
<th>Phytochemical</th>
<th>Bioavailability</th>
</tr>
</thead>
<tbody>
<tr>
<td>Carbohydrates</td>
<td>+++</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+++</td>
</tr>
<tr>
<td>Phenols</td>
<td>+++</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>+++</td>
</tr>
<tr>
<td>Alkaloids</td>
<td>+++</td>
</tr>
<tr>
<td>Tannins</td>
<td>+++</td>
</tr>
<tr>
<td>Saponins</td>
<td>++</td>
</tr>
<tr>
<td>Steroids</td>
<td>+</td>
</tr>
<tr>
<td>Glycosides</td>
<td>ND</td>
</tr>
<tr>
<td>Reducing sugar</td>
<td>ND</td>
</tr>
</tbody>
</table>

Key: + = Present in low concentration; ++ = Present in moderate concentration; +++ = Present in high concentration; ND = Not detected

**MATERIALS AND METHODS**

**Chemicals and reagents/drug**

All chemicals and reagents used for this study were of analytical grade, and were products of Sigma Aldrich, Merck laboratories and Evans Medical plc, Nigeria.

**Plant materials**

The plant material, *Cnidoscolus aconitifolius* leaves were procured from Ihe-Achi in Oji-River L.G.A. of Enugu State of Nigeria. The identification of the plant material was done by the Alfred Ozioko of the Bioresource Development and Conservation Programme (BDCP) Research Centre in Nsukka, Enugu State of Nigeria.

**Preparation of plant materials**

*Cnidoscolus aconitifolius* leaves were washed with tap water and air-dried at room temperature for 21 days. Plant leaves were pulverized using mechanical grinder into coarse form and weighed using electronic weighing balance. A quantity of 500g of the pulsed leaves of *Cnidoscolus aconitifolius* was extracted by maceration in 3 litres of methanol 98% and allowed to stand for 48 hours. The mixture was filtered with mesh followed by Whatman no 1 filter paper. The filtrate was concentrated using rotary evaporator at 40°C to give methanol extract. The extract was stored in the refrigerator at 4°C until needed.

**Phytochemical evaluation**

The screening for the presence of some biologically active constituents of the plant leaf extract was carried out according to the method of Harbone (1973) and Trease and Evans (2012). Quantitative analysis was carried out as described by Harbone (1998), and Soni and Sosa (2013).

**Animal**

Adult male Wistar rats with average weight of 150±11g maintained under suitable conditions (temp 23±2°C, relative humidity 55±10 % and 12 hours light dark cycle) were used for the experiment. The animals were given standard rat pellets and clean water *ad libitum* during the study period. The laboratory animals were used in accordance with laboratory practice regulation and principle of humane laboratory animal care.

**Experimental design**

The rats were randomly divided into 6 groups of 5 rats each. Prior to the experiment, the rats were fasted for 24 hours but allowed free access to water. Group 1 served as normal control, received normal saline (2ml/100g per oral), group 2 received Diclofenac (2ml/100g, p.o) only, group 3 received ranitidine (100mg/kg b.w, p.o) while groups 4 to 6 received methanol extract of *Cnidoscolus aconitifolius* leaves at doses of 100, 200 and 400mg/kg b.w, p.o as a pretreatment. Thirty (30) minutes after pretreatment, diclofenac (150mg/kg b.w) was orally administered to all the rats to induce gastric ulcer. After 6 hours of induction, the rats were sacrificed by inhalation of chloroform in an air tight plastic container and their stomachs were immediately excised. The gastric content was evacuated into test tubes by cutting along the greater curvature of the stomach and centrifuged at 3000 rpm for 10 min for the determination of gastric juice volume, gastric juice pH and the stomach tissues removed and washed with normal saline for determination of gastric ulcer index. Thereafter, a portion of the stomach tissues was taken for histopathological studies.

**Determination of gastric parameters**

Collection of gastric juice: Gastric juice volume was determined using the method described by Kiranmai et al. (2012). The evacuated gastric content was centrifuged at 3000 rpm for 10 min, then separated and the volume measured using a graduated cylinder.

**Determination of pH of the gastric juice**

pH of gastric juice: pH of the gastric juices was determined using a digital pH meter.

**Estimation of gastric ulcerative index changes**

The lesion index was determined by measuring each lesion in mm along its greater length. The stomach tissues opened along greater curvature were washed with normal saline, pinned flat on a cork board and were examined for gastric ulcers with the aid of magnifying lens (×10) and each given a severity rating according to the method of Main and Whittle (1975) as follows: <1mm = 1; > 1mm ≤ 2mm = 2 and >2mm ≤ 3mm = 3. The overall total score divided by 10 was designated as the ulcer index (UI). The percentage ulcer inhibition (percentage ulcer protection) was calculated for each of the groups according to method described by Vinothapooshan and Sundar (2010), using the following equation:
Table 2: Effect of Methanol extract of *Cnidoscolus aconitifolius* leaf on Diclofenac-Induced Gastric lesions in rat

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Volume of Gastric juice (ml)</th>
<th>pH of Gastric juice</th>
<th>Ulcer Index</th>
<th>% Inhibition of Ulcer</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1.24 ±0.17&lt;sup&gt;a&lt;/sup&gt;</td>
<td>8.48 ± 1.25&lt;sup&gt;e&lt;/sup&gt;</td>
<td>0.00±0.00&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-</td>
</tr>
<tr>
<td>Diclofenac only (150 mg/kg b.w)</td>
<td>2.86±0.59&lt;sup&gt;f&lt;/sup&gt;</td>
<td>4.90±0.43&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.48±0.63&lt;sup&gt;e&lt;/sup&gt;</td>
<td>-</td>
</tr>
<tr>
<td>Ranitidine (100 mg/kg b.w)</td>
<td>1.34±0.17&lt;sup&gt;b&lt;/sup&gt;</td>
<td>8.38±2.01&lt;sup&gt;e&lt;/sup&gt;</td>
<td>0.34±0.28&lt;sup&gt;b&lt;/sup&gt;</td>
<td>77.02</td>
</tr>
<tr>
<td>100 mg/kg b.w MECA + Diclofenac</td>
<td>2.12±0.44&lt;sup&gt;e&lt;/sup&gt;</td>
<td>6.12±1.09&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.76±0.80&lt;sup&gt;a&lt;/sup&gt;</td>
<td>48.64</td>
</tr>
<tr>
<td>200 mg/kg b.w MECA + Diclofenac</td>
<td>1.74±0.28&lt;sup&gt;d&lt;/sup&gt;</td>
<td>7.08± 0.55&lt;sup&gt;.e&lt;/sup&gt;</td>
<td>0.72±0.16&lt;sup&gt;e&lt;/sup&gt;</td>
<td>51.35</td>
</tr>
<tr>
<td>400 mg/kg b.w MECA + Diclofenac</td>
<td>1.56±0.48&lt;sup&gt;c&lt;/sup&gt;</td>
<td>7.88±1.40&lt;sup&gt;.e&lt;/sup&gt;</td>
<td>0.36±0.17&lt;sup&gt;b&lt;/sup&gt;</td>
<td>75.68</td>
</tr>
</tbody>
</table>

Values are expressed in mean ± S.D. (n = 5). Values with different alphabets as superscript in the same column differ significantly at p< 0.05.

![Fig. 1](image_url): Macroscopic appearance of gastric mucosa in rats. (a) & (a‘)= Gross and stereomicroscopic view of a normal control rat; (b) & (b‘)= Gross and stereomicroscopic view of a negative ulcer control rat respectively; (c) & (c‘)= Gross and stereomicroscopic view of a rat pre-treated with ranitidine (100mg/kg b.w) respectively; (d) & (d‘)= Gross and stereomicroscopic view of rats pre-treated with 100mg/kg b.w extract respectively; (e) & (e‘)= Gross and stereomicroscopic view of a rat pre-treated with 200mg/kg b.w extract respectively; show minor lesions to the gastric mucosa respectively. (f) & (f‘)= Gross and stereomicroscopic view of a rat pre-treated with 400 mg/kg b.w extract...
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Figure 2: Haematoxylin and eosin stained histological section of gastric ulcer

A: Histological section of a normal control rat: intact gastric pits (black arrow), normal gastric walls and no lesion to the gastric mucosa are observed.
B: Histological section of ulcer negative control rat: extensive mucosal damage, severe loss of surface epithelium and spotty haemorrhagic lesions are seen.
C: Histological section of rat pre-treated with Ranitidine (100 mg/kg b.w): mild mucosal damage, with gastric wall appearance similar to normal.
D: Histological section of a rat pre-treated with 100 mg/kg b.w of extract: mild gastric mucosa lesions, mild loss of epithelium and inflammatory infiltrate, as well as chief cells are seen. There is absence of cellular debris in the lumina.
E: Histological section of a rat pre-treated with 200 mg/kg b.w. of extract showing: typical stomach region with mild granulation of tissue (blue arrow) characterized by the appearance of bumpy thickened structure in the mucosa are visible. The gastric pits are intact with good formation; cellular debris is also found around the superficial zone with numerous degenerate polymorphs (black arrow).
F: Histological section of a rat pre-treated with 400 mg/kg b.w of extract: the intact muscularis mucosa is seen, similar to normal (star).

Percentage protection = \( \frac{(Uc - Ut)}{Uc} \times 100 \)

Uc: Ulcer index in control
Ut: Ulcer index in test

Statistical Analysis
The results obtained were expressed as mean ± SD and were analysed using statistical product and service solutions (SPSS) version 16. Tests of statistical significance were carried out using one-way Analysis of Variance (ANOVA). P values < 0.05 were considered statistically significant.
RESULT

Percentage yield
*Cnidoscolus aconitifolius* leaves yielded 29.94g of the extract which represents 5.99% of the starting plant material.

Phytochemical composition of the methanol extract of *Cnidoscolus aconitifolius* leaf
Preliminary phytochemical screening for some chemical constituents of the extract revealed the presence of soluble carbohydrate, flavonoids, phenols, terpenoids, alkaloids, tannins, steroids and saponins. Reducing sugar and glycosides were not detected as shown in table 1.

Effect of methanol extract of *Cnidoscolus aconitifolius* leaves on gastric juice ph, gastric juice volume gastric ulcer index and percentage ulcer inhibition (percentage ulcer protection) on diclofenac-induced ulcerogenic rats
The gastro protective activity of *Cnidoscolus aconitifolius* leaf extract on diclofenac-induced gastric ulcer in malewistar rats is shown in table 2.

Treatment of rats with diclofenac caused an increase in gastric juice volume and extensive gastric ulcers in the glandular mucosa of stomach, whereas marked decreases in pH of the gastric juice were observed when compared to normal control. Pretreatment of rats with *Cnidoscolus aconitifolius* leaves extract in the doses of 100, 200 and 400mg/kg b.w significantly (p<0.05) suppressed diclofenac induced increases in gastric juice secretion, gastric acidity and formation of gastric lesions in a dose dependent manner when compared to the ulcer control. In rats pre-treated with *Cnidoscolus aconitifolius* leaves extract, the gastro protection of the stomach mucosa were in the range of 48.64 to 75.68% as doses increased from 100 to 400mg/kg b.w. The efficacy of highest tested dose (400mg/kg b.w.) of *Cnidoscolus aconitifolius* leaves extract was similar to ranitidine (100mg/kg b.w.)

Macroscopic examination of diclofenac-induced gastric ulcer in rats
Essentially similar to normal gastric mucosa is presented in pre-treatment groups as shown in fig. 1. Treatment with diclofenac produced gross ulceration and visible black hemorrhagic lesions of gastric mucosa as revealed in the gross (b) and stereomicroscopic (b’) view of ulcer control rats. Gross and stereomicroscopic view of gastric mucosa of rats in pre-treatment groups show remarkable reduction of lesions relative to the ulcer control group while increasing doses of extract, decreased severity of the injuries.

Histopathological evaluation of gastric lesion
The histological study of diclofenac induced gastric mucosa damage in rats is shown in fig. 2.

Administration of diclofenac caused extensive ulceration of gastric mucosa and destruction of the epithelial lining in the group that received diclofenac only. Pretreatment with graded doses of *Cnidoscolus aconitifolius* leaf extract, demonstrated remarkable inhibition of histopathological alterations induced by diclofenac treatment when compared with the ulcer negative control group. At the dose of 400mg/kg b.w., extract and Ranitidine (100mg/kg b.w.) grossly inhibited formation of gastric ulcer ascertained by the appearance of gastric mucosa similar to normal.

DISCUSSION

Pepitic ulcer disease is a chronic inflammatory disease characterized by the ulceration of upper gastrointestinal tract where parietal cells where secrete hydrochloric acid and pepsin (Al-Attar, 2011). However, secretion of gastric acid is central component in the pathogenesis of peptic ulcers (Luiz-Ferreira et al., 2010) by interfering with haemostasis (Green, 1978). More so, gastric acid can inhibit several growth factors that are important for the maintenance of mucosal integrity and for the repair of superficial injury, since these growth factors (e.g. fibroblast growth factor) are acid-labile (Szabo et al., 1994). In general, the properties of NSAIDs that contribute to ulcerezogenesis are categorized into topical irritancy (such as increase in gastrointestinal permeability, decrease in gel hydrophobicity, uncoupling of oxidative phosphorylation in mitochondria etc.) and inhibition of prostaglandin synthesis. In this study, administration of diclofenac produced marked increases in gastric juice volume, and gastric acid secretion (indicated by low pH) in the ulcer control group when compared to normal control as shown in table 2. Also, histological examination of gastric wall of ulcer control group revealed extensive gastric mucosa damage when compared to normal control. This may be attributed to topical irritant properties of diclofenac as well as its ability to inhibit the synthesis of prostaglandin with overall effect of gastro-duodenal ulcers and impairment of mucosal defence, consequently, indicating gastrotoxicity of diclofenac. Pretreatment of rats with ranitidine and graded doses of methanol extract of *Cnidoscolus actinifolius* leaves (100, 200 and 400mg/kg b.w. respectively) significantly (p<0.05) suppressed the diclofenac-induced increases in gastric volume and gastric acid in a dose related manner. Although recent advances have highlighted the multi-factorial pathogenesis of peptic ulcer, secretions of gastric acid is still recognized as a central component of the disease (Luiz-Ferreira et al., 2010).

The antiulcer effect of *Cnidoscolus aconitifolius* could be due to its phytochemical endowment. The presence of phenols and saponins in this plant extract agrees with the findings of Azeez et al. (2010). Flavonoids are a group of
polyphenolic compounds, which exhibit several pharmacological effects in the human body such as anti-inflammatory, anti-hepatotoxic, anti-ulcer, anti-allergic, anti-viral and anti-cancer activities (Uمامaheswari and Sangeetha, 2015). The presence of tannins in the 
Cnidoscolus aconitifolius indicates the astringent properties which facilitate wound healing, good blood circulation and protects the kidney from inflammation (Ross-Ibara and Molina, 2002). The leaf of 
Cnidoscolus aconitifolius contains appreciable amount of phenolics along with other biologically active constituents. The gastro protection observed in this study could be related to the presence of phenolics and flavonoids in the extract which demonstrated antioxidant activities against cellular damage. Barros et al. (2008) reported that phenolic compounds have an antiulcerogenic effect related to cytoprotective activity. Moreover, Kahraman et al. (2003) suggested that flavonoid quercetin promotes a decrease in ulcerative lesions due to its antioxidant effect. In addition, Nwafor et al. (2003) reported that tannins may precipitate 
micro proteins at the side of the ulcer which forms an impervious protective pellicle over the lining to render it less permeable to toxic substances and more resistant to attack of proteolytic enzymes. Also, Lewis and Hansan (1991) and Aguwa and Okunji (1986) reported that triterpenes and saponins possess antiulcer activity through the formation of mucus and inhibitory action on PGF2α. The antioxidant properties of phenols prevent oxidative damage to biomolecules like DNA, lipids, and protein with a specific role in the prevention of chronic diseases such as cancer and cardiovascular diseases (Oyedemi et al., 2012).

CONCLUSION

The result of the present study indicates that oral administration of 
Cnidoscolus aconitifolius leaf extract had a significant and dose dependent inhibition of ulceration. The anti-ulcerogenic effects of the extract could be attributed to its phytoconstituents.

REFERENCES

Cnidoscolus aconitifolius leaf extract protects against hepatic damage induced by chronic ethanol administration in Wistar rats. Alcohol Alcohol, 46: 451-8.

Aguwa CN and Okonji CO (1986). Gastrointestinal studies of 
Pyrenacanthystaudii leaf extracts. J. Ethnopharmacol, 15: 45-55.

Al-Attar MA (2011). Protective effect of 

Awoyinka AO, Balogun IO and Ogunowo AA (2007). Phytochemical screening and in vitro bioactivity of 

Azeez IO, Oyagbemi AA, Oyeyemi MO and Odetola AA (2010). Ameliorative effects of 

antiucler activity of the main phenolic acids found in Brazilian green Propolis. J. Ethnopharmacol., 120: 372-377.


Drury RA, Wallington A and Cameroun SR (1967). In: 

Gastroenterology, 74(1): 38-43.


Toxicology, 183: 133-42.

Kiranmai M, Sri BU, Reddy DS, Kumar CBM and Ibrahim M (2012). Evaluation of total phenolic 
contents and antiulcerogenic activity of root bark of 

Lewis DA and Hanson D (1991). Anti-ulcer drugs of 

Luiz-Ferreira A, De Ameida ACA and Cola M (2010). 
Mechanisms of the gastric antiulcerogenic activity of 
Anarcadiumhumile on ethanol-induced acute gastric mucosal injury in rats. 

Main IH and Whittle BJ (1975). Investigation of the 
vasodilator and antiserotory role of prostaglandins in the 
rat gastric mucosa by use of non-steroidal anti-inflammatory drugs. 

Annon muricata


