Impairment of liver synthetic function and the production of plasma proteins in primary breast cancer patients on doxorubicin-cyclophosphamide (AC) protocol

Zikria Saleem¹, Mobasher Ahmad¹, Furqan Khurshid Hashmi¹, Hamid Saeed¹* and Muhammad Tahir Aziz²

¹University College of Pharmacy, University of the Punjab, Allama Igbal Campus, Lahore, Pakistan.

Abstract: Doxorubicin and Cyclophosphamide (AC protocol) combination is usually considered as a first line therapy in newly diagnosed breast cancer patients. Thus, a retrospective observational study was conducted to monitor the effect of AC protocol on liver synthetic functions and production of plasma proteins in breast cancer patients, reporting to specialized cancer care hospital of Lahore, Pakistan. A total of 75 patients (n=75) on AC protocol with breast cancer were observed in this study. The patient data including age, gender, body surface area, dosage, disease status and laboratory biochemical values were recorded by reviewing historical treatment records. Pre-treatment values were taken as baseline values for albumin, globulin, blood urea nitrogen (BUN), albumin/globulin (A/G) ratio and total proteins. The baseline values were compared after each cycle of by applying ANOVA using statistical tool SPSS® version 21. The plasma levels of blood urea nitrogen (BUN), total protein and globulin dropped significantly (p<0.05) in patients of all age groups. However, the albumin levels were not significantly changed (p>0.05). The A/G ratio level increased (p<0.05) as a result of reduction in globulin levels. Significant changes in plasma protein levels were observed in the elderly patients (50 to 65 years) than patients between 20 to 50 years of age. AC protocol impairs liver synthetic functions as observed by decreased blood urea nitrogen (BUN) and plasma protein levels.

Keywords:Doxorubicin-Cyclophosphamide, primary breast cancer, hepatic proteins, blood urea nitrogen (BUN).

INTRODUCTION

Liver is a pivotal organ of the body because it is involved in the regulation of vital functions including biosynthesis, biotransformation and excretion of foreign chemicals (Pavek and Dvorak, 2008). One of its crucial function is to synthesize important proteins which are involved in normal functioning of the body (JE, 2011). Main proteins synthesized by hepatocytes are albumin and globulin. Albumin is the most abundant protein found in the extra cellular fluid (EFC) and accounts for 60% of the total protein in plasma (Turell et al., 2013). Additionally, albumin is the most valuable prognostic and diagnostic laboratory marker of liver synthetic functions (Doweiko and Nompleggi, 1991). Major function of albumin is to bind various physiological compounds including acidic drugs (Shargel et al., 2005). Albumin also regulates colloidal osmotic pressure of the blood (Boldt, 2010). Globulin proteins, synthesized from liver, are divided into α 1-globulins and β -globulins while γ -globulin is synthesized in lymphocytes (Murray et al., 2009, Miller et al., 1951). Beside binding basic drugs, these proteins regulate transport of cholesterol and iron as well as play their structural role in forming mucoproteins, plasminogens, prothrombin and other clotting factors (Murray et al., 2009). Decreased level of these proteins is used as a hallmark to diagnose liver disorders (Hauser et

al., 1996). It has been shown by number of studies that the liver diseases affecting liver synthetic functions could contribute to hypoalbuminaemia, low total protein and a change in A/G ratio(Dubois et al., 2006, Throop et al., 2004). Likewise, impaired protein synthesis by the liver could lead to diverse clinical manifestations such as, coagulopatheies, disseminated intravascular coagulopathy (DIC), increased blood cholesterol concentrations, ascites and peripheral edema (McPherson and Pincus, 2011). Moreover, a decrease in plasma protein levels has been suggested to increase the free plasma drug concentration (unbound drug) - leading to increase volume of distribution of the drug and significantly higher probability of unwanted toxic effects (Rolan, 1994, Blaschke, 1977). Liver also play its role in converting toxic ammonia molecules into urea which can easily be excreted through kidney (Landsberg et al., 2005).

Literature evidences suggest a close link between an organ damage and chemotherapy. (Hashmi *et al.*, 2012). Drug-induced hepato-toxicity is mainly due to direct involvement of liver in the metabolism of foreign chemicals, including drugs (Maurel and Rosenbaum, 2012, Holt and Ju, 2006). AC protocol is used as a first line therapy in the treatment of newly diagnosed patients with breast cancer (Evans *et al.*, 2005, Fisher *et al.*, 1990). Doxorubicin, an anthracycline antibiotic, can cause subclinical hepatotoxicity in patients during course of

²Shaukat Khanum Cancer Hospital and Research Center, Lahore, Pakistan

^{*}Corresponding author: e-mail: hamid.pharmacy@pu.edu.pk

therapy (Rashid et al., 2013, Injac et al., 2008, Klasco, 2009). Hepatotoxic effect of doxorubicin is more pronounced in the cancer patients using doxorubicin in combination therapy (Kalender et al., 2005). Out of 71% total serum binding (Chassany et al., 1996), 62% of doxorubicin bind to albumin (Eksborg et al., 1982). Cyclophosphamide, an alkylating agent, is a pro-drug activated by hepatic cytochromal P450 enzyme system (Chen et al., 2004). Cyclophosphamide minimally binds to serum proteins, however, its metabolites bind considerably (>60%) to serum proteins (Grochow and Colvin, 1979). Cyclophosphamide has also been shown to cause hepatotoxicity with altered liver synthetic functions (Subramaniam et al., 2013), owing to significant oxidative stress and generation of free radicals (Catimel et al., 1994, Meyer et al., 2012). During the course of cancer treatment many patients die owing to severity of the disease, nevertheless, many of these are related to anticancer therapy (Swerdlow et al., 2007), thus warrant regular monitoring (Galpin and Evans, 1993, Steinherz et al., 1992). These literature evidences suggest that among others, effect of anti-cancer therapy on liver synthetic functions pose significant risk of drug induced hepatic toxicity in cancer patients. However, little attention is paid towards the effect of AC protocol on hepatic synthetic functions during anticancer therapy. In this study, we evaluated the overall production of plasma proteins, albumin globulin and blood urea nitrogen (BUN) in breast cancer patients under AC protocol. Interestingly, total plasma protein, globulin and blood urea nitrogen (BUN) levels were significantly reduced in all patients undergoing anti-cancer therapy. However, in the elderly patients (51 to 65 years) total plasma protein, globulin and blood urea nitrogen (BUN) reduction was more significant compared to patients between 20 to 50 years of age.

Aim of the study

The aim of the study was to assess the liver synthetic functions focusing blood urea nitrogen (BUN) and protein synthesis - total, albumin and globulin protein levels, in breast cancer patients of different age groups

METHODS

Study design

A retrospective observational study was designed to see the effect of AC protocol on liver synthetic functions in synthesizing liver plasma proteins and urea. Patients on AC-protocol for primary breast cancer were selected. The patient's medical records were checked to evaluate changes in the laboratory values during course of therapy. Pre-treatment baseline values of albumin, globulin, albumin/globulin (A/G) ratio, blood urea nitrogen (BUN) and total protein were checked before initiating and after drug therapy during 4 different cycles (cycle 1, cycle 2, cycle 3 and cycle 4) with the span of 21 days. This

retrospective study was designed to check the correlation between AC chemotherapy protocol and its effect on liver protein synthesis.

Data source

Data was collected from specialized cancer care hospital in Lahore Pakistan after the approval of ethical committees of the hospital and University College of Pharmacy, University of the Punjab, Lahore, Pakistan, reference number EC/UCP/092/2013.

Sample size and distribution

Total population size of this study was 75 patients (n=75). Out of this total population, 18 patients (24%) were in age group 20-35 years of age, 40 patients (53%) were in age group 36-50 years of age and 17 (23%) patients were in age group 51-65 years of age.

Inclusion criteria

The inclusion criteria were as follows; a) female patients diagnosed with primary breast cancer, b) patients treated with doxorubicin and cyclophosphamide. c) patients administered with 4 cycles of chemotherapy agents repeated after every 21 days, d) Patients from 20 to 65 years of age, e) patients with ECOG (eastern cooperative oncology group) performance status 0-2 were selected (Sørensen *et al.*, 1993), f) history of no clinical hepatic disease.

STATISTICAL ANALYSIS

Data was analyzed by using SPSS version 21.0. All the parameters were noted as standard deviation of the mean. The baseline values were compared after each cycle of doxorubicin-cyclophosphamide (AC) cycle by applying ANOVA using statistical tool SPSS version 21. A statistical value of $p \le 0.05$ was considered significant.

RESULTS

Chemotherapeutic agents usually exhibit narrow therapeutic index – making tissues and organs susceptible to chemotherapeutic toxicity (Alnaim, 2007). Thus, we aimed at examining the effect of AC protocol on liver's ability to synthesize proteins, i.e., albumin and globulin. As shown in table. 1 average baseline profiles for total protein, albumin and globulin remain unchanged in combined age group and in split age groups intended in an ascending order (table 1 & fig. 1B-C). Similar trend was observed for albumin/globulin ratio (A/G ratio) and blood urea nitrogen (BUN) (table 1). However, compared to baseline, mild reduction in total protein (4%) and marked reduction in globulin levels (8%) were observed in all age groups with every programmed cycle, nevertheless, intracycle differences were negligible (table 2). However, there is highly significant reduction (21%) in the level of blood urea nitrogen (BUN) after completion of therapy in

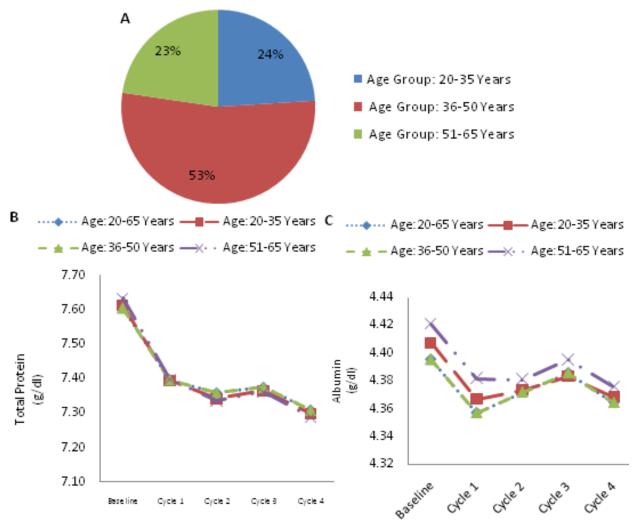


Fig. 1A): Pei chart showing percentage age distribution of female breast cancer patients. **B**) Macro-graph showing total plasma protein levels in different age groups at baseline and after every programmed AC chemotherapy cycle. **C**) Macro-graph showing plasma albumin protein levels in different age groups at baseline and after every programmed AC chemotherapy cycle.

total population (tables 1 & 4). Strikingly, compared to baseline, albumin levels were not significantly affected by AC protocol with every programmed cycle in all age groups (tables 1 & 3).

Aging is a progressive, consistent and time related loss of body's functional units (Crooks *et al.*, 1976, Turnheim, 2003, Mangoni and Jackson, 2004). Age related decline in the concentration of plasma protein levels, albumin and α-1 glycoproteins, have not been reported (Fu and Nair, 1998). Additionally, there is scanty of literature evidences that point towards unwanted effects of AC protocol on plasma albumin and globulin levels. Our data suggest that breast cancer is more prevalent in females in 4 and 5th decade of life (36-50 yrs) as compared to other age groups (fig. 1A). Interestingly, compared to baseline values, the effect of AC protocol was more pronounced on aged patient's (51-65 yrs) globulin levels (p<0.001), while there were no intra-cycle differences, in globulins levels,

in younger patients compared to significant reduction, after cycle 4, in aged patients (9%) (tables 1 & 2, fig 2A). Similarly, compared to baseline A/G ratios, the A/G ratios were higher in aged patients (p≤0.003) than younger patients after cycle 4 therapy, while intra-cycle A/G ratios remained unchanged (table. 3 & fig. 2 B). The average baseline value of blood urea nitrogen (BUN) in patients of age group 51-65 years was 11.29mg/dl and declined to 8.65 mg/dl after cycle 4. It showed that it was the most significant reduction (23%) in the level of blood urea nitrogen (BUN) in elderly patients (tables 1 & 4, fig. 2C).

DISCUSSION

For women, with metastatic breast cancer, the median survival range is within or about 2 years. Therefore, a number of drug regimens have been employed, such as, AC (doxorubicin and cyclophosphamide), CMF (cyclophosphamide, methotrexate and 5-flurouracil), CAF

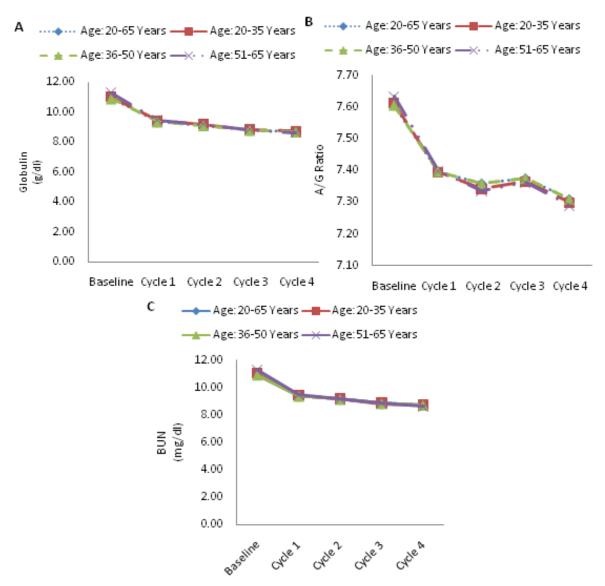


Fig. 2A: Macro-graph showing plasma globulin levels at baseline and after every programmed AC chemotherapy cycles. B) A/G ratio, calculated from plasma albumin and globulin values, at baseline and after each chemotherapy cycles. C) Macro-graph showing blood urea nitrogen (BUN) levels at baseline and after every programmed AC chemotherapy cycles.

(doxorubicin, cyclophosphamide and 5-flurouracil) and doxorubicin plus paclitaxel combination, as a first line chemotherapy in breast cancer (Tranum *et al.*, 1982, Buzdar *et al.*, 1989, Jassem *et al.*, 2001). Despite clinical benefits, in form of prolong survival to breast cancer patients, these drug regimens are prone to cause organ/tissue toxicity, thus, beg further investigations. The frequency of breast cancer in developing countries is lower compared to western countries and is attributed to younger population pyramid (Bibhusal Thapa *et al.*, 2013). However, recent trend seems alarming, suggesting that the frequency of breast cancer is increasing in younger females in Asia region, including Pakistan (Yoo, 2010). Our data point towards the similar trend showing significant breast cancer prevalence in relatively younger

(18%) and middle age groups (53%) rather than older patients (17%).

In this study we focused on the effect of AC chemotherapy protocol on total plasma protein, plasma albumin, globulin and urea levels produced by liver. Data suggest that AC (Doxorubicin and Cyclophosphamide) chemotherapy protocol significantly affected the total plasma proteins, globulin and blood urea nitrogen (BUN) levels in all age groups with more significant reduction in globulin protein and blood urea nitrogen (BUN) levels in elderly patients, in 5th and 6th decade of life. However, no changes in albumin protein levels were observed. It is beyond doubt that liver plays an important role in protein and drug metabolism (Barle *et al.*, 1997) and profound

Table 1: Mean baseline and post therapy values of plasma proteins in different age group

Lab Values	Age (Years)	Baseline Value	After Cycle 1	After Cycle 2	After Cycle 3	After Cycle 4
Lab values	20-65	7.60±0.470	7.40±0.469	7.36±0.412	7.38±0.390	7.31±0.370
Total Protein	20-35	7.60 ± 0.470 7.61 ± 0.481	7.40 ± 0.390 7.40 ± 0.390	7.30 ± 0.412 7.34 ± 0.350	7.38 ± 0.390 7.37 ± 0.410	7.31 ± 0.370 7.30 ± 0.395
(g/dl)	36-50	7.60 ± 0.481 7.60 ± 0.510	7.40 ± 0.390 7.40 ± 0.470	7.34±0.330 7.36±0.380	7.38 ± 0.410 7.38 ± 0.425	7.30 ± 0.393 7.31 ± 0.405
(g/ui)	51-65	7.63 ± 0.525	7.40 ± 0.470 7.40 ± 0.495	7.33±0.470	7.36 ± 0.423 7.36 ± 0.412	7.31 ± 0.403 7.29 ± 0.350
	20-65	4.40 ± 0.319	4.36 ± 0.327	4.37 ± 0.315	4.39 ± 0.319	4.36 ± 0.313
A 11	20-03	4.40 ± 0.319 4.41 ± 0.325	4.30 ± 0.327 4.37 ± 0.319	4.37 ± 0.313 4.37 ± 0.316	4.39 ± 0.319 4.38 ± 0.318	4.30 ± 0.313 4.37 ± 0.320
Albumin		_			_	
(g/dl)	36-50	4.40±0.318	4.36±0.312	4.37 <u>±</u> 0.310	4.39±0.319	4.36±0.315
	51-65	4.42±0.322	4.38±0.319	4.38±0.318	4.40 <u>±</u> 0.319	4.38±0.314
a:	20-65	3.20±0.296	3.04 <u>±</u> 0.286	3.00 <u>±</u> 0.278	3.00 <u>±</u> 0.286	2.95±0.296
Globulin	20-35	3.20 <u>±</u> 0.231	3.03 <u>±</u> 0.296	2.98 <u>±</u> 0.276	2.98 <u>±</u> 0.299	2.94 <u>±</u> 0.291
(g/dl)	36-50	3.20 <u>±</u> 0.296	3.04 <u>±</u> 0.279	3.00 <u>±</u> 0.281	3.00 <u>±</u> 0.296	2.95 <u>±</u> 0.231
	51-65	3.20 <u>±</u> 0.233	3.02 <u>±</u> 0.296	2.97 <u>±</u> 0.231	2.96 <u>±</u> 0.289	2.91 <u>±</u> 0.296
	20-65	1.39 <u>±</u> 0.148	1.45 <u>±</u> 0.158	1.48 <u>±</u> 0.161	1.48 <u>±</u> 0.158	1.49 <u>±</u> 0.158
A/G Ratio	20-35	1.39 <u>±</u> 0.155	1.45 <u>±</u> 0.159	1.48 <u>±</u> 0.159	1.49 <u>±</u> 0.158	1.50 <u>±</u> 0.161
A/G Ratio	36-50	1.39 <u>±</u> 0.158	1.45 <u>±</u> 0.163	1.48 <u>±</u> 0.153	1.48 <u>±</u> 0.151	1.49 <u>±</u> 0.158
	51-65	1.40 <u>±</u> 0.149	1.47 <u>±</u> 0.158	1.50 <u>±</u> 0.157	1.50 <u>±</u> 0.156	1.52 <u>±</u> 0.162
	20-65	10.92 <u>±</u> 0.45	9.35 <u>±</u> 0.40	9.12 <u>±</u> 0.39	8.80 <u>+</u> 0.37	8.68 <u>±</u> 0.35
BUN	20-35	11.03 <u>±</u> 0.43	9.42 <u>±</u> 0.41	9.20 <u>±</u> 0.38	8.84 <u>±</u> 0.38	8.74 <u>±</u> 0.39
(mg/dl)	36-50	10.92 <u>±</u> 0.47	9.35 <u>±</u> 0.39	9.12 <u>±</u> 0.39	8.80 <u>±</u> 0.39	8.68 <u>±</u> 035
	51-65	11.29 <u>±</u> 0.44	9.48 <u>±</u> 0.39	9.21 <u>±</u> 0.37	8.85 <u>±</u> 0.40	8.65 <u>±</u> 0.38
Lab Values	Age (Years)	Baseline Value	After Cycle 1	After Cycle 2	After Cycle 3	After Cycle 4
	20-65	7.60 <u>±</u> 0.470	7.40 <u>±</u> 0.469	7.36 <u>±</u> 0.412	7.38 <u>±</u> 0.390	7.31 <u>±</u> 0.370
Total Protein	20-35	7.61 <u>±</u> 0.481	7.40 <u>±</u> 0.390	7.34 <u>±</u> 0.350	7.37 <u>±</u> 0.410	7.30 <u>±</u> 0.395
(g/dl)	36-50	7.60 <u>±</u> 0.510	7.40 <u>±</u> 0.470	7.36 <u>±</u> 0.380	7.38 <u>±</u> 0.425	7.31 <u>±</u> 0.405
	51-65	7.63 <u>±</u> 0.525	7.40 <u>±</u> 0.495	7.33 <u>±</u> 0.470	7.36 <u>±</u> 0.412	7.29 ± 0.350
	20-65	4.40 <u>±</u> 0.319	4.36 <u>±</u> 0.327	4.37 <u>±</u> 0.315	4.39 <u>±</u> 0.319	4.36 <u>±</u> 0.313
Albumin	20-35	4.41 <u>±</u> 0.325	4.37 <u>±</u> 0.319	4.37 <u>±</u> 0.316	4.38 <u>±</u> 0.318	4.37 <u>±</u> 0.320
(g/dl)	36-50	4.40 <u>±</u> 0.318	4.36±0.312	4.37 <u>±</u> 0.310	4.39 <u>±</u> 0.319	4.36±0.315
,	51-65	4.42 ± 0.322	4.38 <u>±</u> 0.319	4.38 <u>±</u> 0.318	4.40 <u>±</u> 0.319	4.38±0.314
	20-65	3.20±0.296	3.04 <u>±</u> 0.286	3.00 <u>±</u> 0.278	3.00 <u>±</u> 0.286	2.95 ± 0.296
Globulin	20-35	3.20±0.231	3.03±0.296	2.98 <u>±</u> 0.276	2.98±0.299	2.94±0.291
(g/dl)	36-50	3.20±0.296	3.04 <u>±</u> 0.279	3.00 <u>±</u> 0.281	3.00±0.296	2.95±0.231
(C)	51-65	3.20 <u>±</u> 0.233	3.02 <u>±</u> 0.296	2.97 <u>±</u> 0.231	2.96 <u>±</u> 0.289	2.91±0.296
						1.49 ± 0.158
	20-65	1.39 <u>±</u> 0.148	1.45 <u>±</u> 0.158	1.48 <u>±</u> 0.161	1.48 <u>±</u> 0.158	1.49 <u>±</u> 0.158 1.50±0.161
A/G Ratio	20-65 20-35	1.39 <u>±</u> 0.148 1.39 <u>±</u> 0.155	1.45 <u>±</u> 0.158 1.45 <u>±</u> 0.159	1.48±0.161 1.48±0.159	1.48±0.158 1.49±0.158	1.50 <u>±</u> 0.161
A/G Ratio	20-65 20-35 36-50	1.39±0.148 1.39±0.155 1.39±0.158	1.45±0.158 1.45±0.159 1.45±0.163	1.48±0.161 1.48±0.159 1.48±0.153	1.48±0.158 1.49±0.158 1.48±0.151	1.50 <u>±</u> 0.161 1.49 <u>±</u> 0.158
A/G Ratio	20-65 20-35 36-50 51-65	1.39±0.148 1.39±0.155 1.39±0.158 1.40±0.149	1.45±0.158 1.45±0.159 1.45±0.163 1.47±0.158	1.48±0.161 1.48±0.159 1.48±0.153 1.50±0.157	$\begin{array}{c} 1.48 \pm 0.158 \\ 1.49 \pm 0.158 \\ 1.48 \pm 0.151 \\ 1.50 \pm 0.156 \end{array}$	1.50±0.161 1.49±0.158 1.52±0.162
	20-65 20-35 36-50 51-65 20-65	1.39±0.148 1.39±0.155 1.39±0.158 1.40±0.149 10.92±0.45	$\begin{array}{c} 1.45 \pm 0.158 \\ 1.45 \pm 0.159 \\ 1.45 \pm 0.163 \\ 1.47 \pm 0.158 \\ 9.35 \pm 0.40 \end{array}$	$\begin{array}{c} 1.48 \pm 0.161 \\ 1.48 \pm 0.159 \\ 1.48 \pm 0.153 \\ 1.50 \pm 0.157 \\ 9.12 \pm 0.39 \end{array}$	$\begin{array}{c} 1.48 \pm 0.158 \\ 1.49 \pm 0.158 \\ 1.48 \pm 0.151 \\ 1.50 \pm 0.156 \\ 8.80 \pm 0.37 \end{array}$	1.50±0.161 1.49±0.158 1.52±0.162 8.68±0.35
BUN	20-65 20-35 36-50 51-65 20-65 20-35	$ \begin{array}{c} 1.39 \pm 0.148 \\ 1.39 \pm 0.155 \\ 1.39 \pm 0.158 \\ 1.40 \pm 0.149 \\ 10.92 \pm 0.45 \\ 11.03 \pm 0.43 \end{array} $	1.45±0.158 1.45±0.159 1.45±0.163 1.47±0.158 9.35±0.40 9.42±0.41	1.48±0.161 1.48±0.159 1.48±0.153 1.50±0.157 9.12±0.39 9.20±0.38	1.48±0.158 1.49±0.158 1.48±0.151 1.50±0.156 8.80±0.37 8.84±0.38	1.50±0.161 1.49±0.158 1.52±0.162 8.68±0.35 8.74±0.39
	20-65 20-35 36-50 51-65 20-65	1.39±0.148 1.39±0.155 1.39±0.158 1.40±0.149 10.92±0.45	$\begin{array}{c} 1.45 \pm 0.158 \\ 1.45 \pm 0.159 \\ 1.45 \pm 0.163 \\ 1.47 \pm 0.158 \\ 9.35 \pm 0.40 \end{array}$	$\begin{array}{c} 1.48 \pm 0.161 \\ 1.48 \pm 0.159 \\ 1.48 \pm 0.153 \\ 1.50 \pm 0.157 \\ 9.12 \pm 0.39 \end{array}$	$\begin{array}{c} 1.48 \pm 0.158 \\ 1.49 \pm 0.158 \\ 1.48 \pm 0.151 \\ 1.50 \pm 0.156 \\ 8.80 \pm 0.37 \end{array}$	1.50±0.161 1.49±0.158 1.52±0.162 8.68±0.35

changes in the rate of protein synthesis by the liver can occur in response to hepatic insult, affecting drug protein binding and disposition (LEVELS, 2000, Sallie *et al.*, 1991, Wolfe *et al.*, 1989). Furthermore, according to research literature, concurrent administration of anticancer drugs can induce oxidative stress in addition to drug metabolic workload on the liver maximizing hepatotoxic prospects (Pieniążek *et al.*, 2013). Thus, routinely, the most common biomarker employed for liver function test (LFT) include, ALT and albumin nevertheless, inclusion of other proteins and markers, such as total

globulins (LeBlond *et al.*, 2004) and A/G ratio could also provide a mean to assess liver synthetic functions (Pratt and Kaplan, 2000, El–Serag and Rudolph, 2007). In our study, we found marked reduction in the total plasma protein and globulin levels in all age groups – compared to baseline values, however, no change in albumin levels was observed. There is considerable controversy regarding the inclusion of total protein in liver function test, despite the fact that 73% of hospital laboratories in USA and UK use total protein in liver function profiles

Table 2:Multiple comparison of total protein and globulin levels between baseline and every chemotherapy cycle and intra-cycle comparison

Multiple Comparisons		Total Protein			Globulin		
Cycles (I)	Cycles (J)	Mean Difference (I-J)	Std. Error	Sig.	Mean Difference (I-J)	Std. Error	Sig.
(Cycle1	.20711*	.06887	.023	.16267*	.05220	.017
	Cycle2	.24493*	.06887	.004	.20467*	.05220	.001
Baseline	Cycle3	.22907*	.06887	.009	.20453*	.05220	.001
	Cycle4	.29467*	.06887	.000	.24920*	.05220	.000
	Cycle2	.03783	.06887	.982	.04200	.05220	.929
Cycle1	Cycle3	.02196	.06887	.998	.04187	.05220	.930
	Cycle4	.08756	.06887	.709	.08653	.05220	.462
C1-2	Cycle3	01587	.06887	.999	00013	.05220	1.000
Cycle2	Cycle4	.04973	.06887	.951	.04453	.05220	.914
Cycle3	Cycle4	.06560	.06887	.876	.04467	.05220	.913

Table 3: Multiple Comparison of albumin and A/G ratio between baseline and every programmed chemotherapy cycles and among chemotherapy cycles

Multiple Comparisons		Total Protein			Globulin		
Cycles (I)	Cycles (J)	Mean Difference (I-J)	Std. Error	Sig.	Mean Difference (I-J)	Std. Error	Sig.
Baseline	Cycle1	.20711*	.06887	.023	.16267*	.05220	.017
	Cycle2	.24493*	.06887	.004	.20467*	.05220	.001
	Cycle3	.22907*	.06887	.009	.20453*	.05220	.001
	Cycle4	.29467*	.06887	.000	.24920*	.05220	.000
	Cycle2	.03783	.06887	.982	.04200	.05220	.929
Cycle1	Cycle3	.02196	.06887	.998	.04187	.05220	.930
	Cycle4	.08756	.06887	.709	.08653	.05220	.462
Constant	Cycle3	01587	.06887	.999	00013	.05220	1.000
Cycle2	Cycle4	.04973	.06887	.951	.04453	.05220	.914
Cycle3	Cycle4	.06560	.06887	.876	.04467	.05220	.913

(Hayden and van Heyningen, 2001). Aging has been associated with decrease serum albumin levels with no change in total globulin levels in females irrespective of menstrual function (Sokoll and Dawson-Hughes, 1989). The unchanged serum albumin levels in our study could be attributed to prolong half life of albumin (20 days) and re-synthesis plus normalization of albumin in 14 days, therefore, measuring serum albumin levels after every 21 days of chemotherapy cycle is uninformative (Beck and Rosenthal, 2002). However, unchanged albumin also suggests normal drug protein binding, especially for doxorubicin (25). Reduced serum albumin levels in cancer patients, have been associated with poor survival while its normal or higher values serve as a powerful prognostic variable and are associated with patient survival time (Lis et al., 2003). However, reduced serum total globulin levels in all age groups point towards the immune-suppressive effect of AC chemotherapy protocol in breast cancer patients and could be employed as an important supplementary parameter of prognostic value in advanced stage breast cancer patients (Fatima et al., 2013, Al-Joudi, 2005). Besides, more significant changes with aging could be attributed towards an age dependent decline in immune utility, in aged cancer patients susceptible to cyto-toxic chemotherapeutic agents because

of the decrease hepatic blood flow and reduce drug clearance through kidney (Klotz, 2009, Shi and Klotz, 2011, Corsonello et al., 2010, Triantafyllou et al., 2010, Aapro et al., 2011). We also observed significant increase in A/G ratio (7.46 %) in all age groups, compared to base line values with more pronounced changes after cycle 4. According to literature evidences, increased A/G ratio could be associated with improved survival in patients with normal albumin levels and normally higher A/G ratio is observed in pre-treated breast cancer patients with lower albumin levels and higher globulins levels a compensatory mechanism to counter albumin reduction and to further improve immune competence (Azab et al., 2013). Decline in urea levels indicate an incomplete conversion of ammonia into urea. So as a result of impair liver synthetic function to synthesize urea through urea cycle, severe hyperammonemia induced hepatic encephalopathy can further worsen the condition of the patient (Ong et al., 2003).

In conclusion, our data suggest that AC protocol opted for breast cancer patients affect total plasma protein, blood urea nitrogen (BUN) and globulin levels, in all age groups with more significant effect in the elderly globulin levels, after every programmed chemotherapy cycle. However,

Multiple Comparisons	BUN			
Cycles (I)	Cycles (J)	Mean Difference (I-J)	Std. Error	Sig.
	Cycle1	1.56227*	.47395	.009
Danalina	Cycle2	1.79867*	.47395	.002
Baseline	Cycle3	2.11307*	.47395	.000
	Cycle4	2.23080^*	.47395	.000
	Cycle2	.23640	.47395	.987
Cycle1	Cycle3	.55080	.47395	.773
-	Cycle4	.66853	.47395	.621
Cycle2	Cycle3	.31440	.47395	.964
Cycle2	Cycle4	.43213	.47395	.892
Cycle3	Cycle4	11773	47395	999

Table 4:Multiple Comparison of BUN between baseline and every programmed chemotherapy cycles and among chemotherapy cycles

no changes in albumin levels were observed due to albumin physiological and biochemical regulation. Moreover, confirming other reports, our data suggest profound prevalence of breast cancer in younger and middle age patients. Decreased globulin levels with normal albumin levels supporting increased A/G ratio, could possibly predict better survival and long term mortality in breast cancer patients, nonetheless, further studies are required to further understand the concept.

REFERENCES

- Aapro M, Bernard-Marty C, Brain E, Batist G, Erdkamp F, Krzemieniecki K, Leonard R, Lluch A, Monfardini S and Ryberg M (2011). Anthracycline cardiotoxicity in the elderly cancer patient: A SIOG expert position paper. *Annals of Oncology*, **22**: 257-267.
- AL-Joudi F (2005). Prognostic value of an index for serum globulin compensation in colon and breast cancers. *Singapore medical journal*, **46**: 710.
- Alnaim L (2007). Therapeutic drug monitoring of cancer chemotherapy. *J Oncol Pharm Pract*, **13**: 207-221.
- Azab BN, Bhatt VR, Vonfrolio S, Bachir R, Rubinshteyn V, Alkaied H, Habeshy A, Patel J, Picon AI & Bloom SW (2013). Value of the pretreatment albumin to globulin ratio in predicting long-term mortality in breast cancer patients. *Am J Surg*, **206**: 764-770.
- Barle H, Nyberg B, Essen P, Andersson K, Mcnurlan MA, Wernerman J and Garlick PJ (1997). The synthesis rates of total liver protein and plasma albumin determined simultaneously *in vivo* in humans. *Hepatology*, **25**: 154-158.
- Beck FK and Rosenthal TC (2002). Prealbumin: A marker for nutritional evaluation. *Am Fam Physician*, 65.
- Bibhusal Thapa YS, Sayami P, Shrestha UK, Sapkota R and Sayami G (2013). Breast cancer in young women from a low risk population in nepal. *Asian Pac J Cancer Prev*, **14**: 5095-5099.
- Blaschke TF (1977). Protein binding and kinetics of drugs in liver diseases. *Clin Pharmacokinet*, **2**: 32-44.

- Boldt J (2010). Use of albumin: An update. *Brit J Anaesth*, **104**: 276-284.
- Buzdar AU, Kau SW, Smith TL & Hortobagyi GN (1989). Ten-year results of FAC adjuvant chemotherapy trial in breast cancer. *Am J Clin Oncol*, **12**: 123-128.
- Catimel G, Chauvin F, Guastalla J, Rebattu P, Biron P and Clavelt M (1994). FAC (fluorouracil, doxorubicin, cyclophosphamide) as second line chemotherapy in patients with metastatic breast cancer progressing under FEC (fluorouracil, epirubicin, cyclophosphamide) chemotherapy. *Ann Oncol*, **5**: 95-97.
- Chassany O, Urien S, Claudepierre P, Bastian G & Tillement JP (1996). Comparative serum protein binding of anthracycline derivatives. *Cancer Chemoth Pharm*, **38**: 571-573.
- Chen CS, Lin JT, Goss KA, He YA, Halpert JR and Waxman DJ (2004). Activation of the anticancer prodrugs cyclophosphamide and ifosfamide: Identification of cytochrome P450 2B enzymes and site-specific mutants with improved enzyme kinetics. *Mol Pharmacol*, **65**: 1278-1285.
- Corsonello A, Pedone C and Incalzi RA (2010). Agerelated pharmacokinetic and pharmacodynamic changes and related risk of adverse drug reactions. *Curr Med Chem.* 17: 571-584.
- Crooks J, O'malley K and Stevenson I (1976). Pharmacokinetics in the elderly. *Clin Pharmacokinet*, 1: 280-296.
- Doweiko JP and Nompleggi DJ (1991). Reviews: Role of albumin in human physiology and patho physiology. *J Parenter Enteral Nutr*, **15**: 207-211.
- Dubois MJ, Orellana-Jimenez C, Melot C, De Backer D, Berre J, Leeman M, Brimioulle S, Appoloni O, Creteur J and Vincent JL (2006). Albumin administration improves organ function in critically ill hypoalbuminemic patients: A prospective, randomized, controlled, pilot study*. *Crit Care Med*, 34: 2536-2540.
- Eksborg S, Ehrsson H and Ekqvist B (1982). Protein binding of anthraquinone glycosides, with special reference to adriamycin. *Cancer Chemoth Pharm*, **10**: 7-10.

- El-Serag HB and Rudolph KL (2007). Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. *Gastroenterology*, **132**: 2557-2576.
- Evans TJ, Yellowlees A, Foster E, Earl H, Cameron DA, Hutcheon AW, Coleman RE, Perren T, Gallagher CJ and Quigley M (2005). Phase III randomized trial of doxorubicin and docetaxel versus doxorubicin and cyclophosphamide as primary medical therapy in women with breast cancer: An anglo-celtic cooperative oncology group study. *J Clin Oncol*, 23: 2988-2995.
- Fatima T, Roohi N and Abid R (2013). Circulatory Proteins in women with breast cancer and their chemotherapeutic responses. *Pakistan Journal Of Zoology*, **45**: 1207-1213.
- Fisher B, Brown AM, Dimitrov NV, Poisson R, Redmond C, Margolese RG, Bowman D, Wolmark N, Wickerham DL and Kardinal CG (1990). Two months of doxorubicin-cyclophosphamide with and without interval reinduction therapy compared with 6 months of cyclophosphamide, methotrexate and fluorouracil in positive-node breast cancer patients with tamoxifennonresponsive tumors: Results from the national surgical adjuvant breast and bowel project B-15. *J Clin Oncol*, 8: 1483-1496.
- Fu A and Nair KS (1998). Age effect on fibrinogen and albumin synthesis in humans. *Am J Physio-Endocrinol Metab*, **275**: E1023-E1030.
- Galpin AJ and Evans WE (1993). Therapeutic drug monitoring in cancer management. *Clinical Chemistry*, **39**: 2419-2430.
- Grochow LB and Colvin M (1979). Clinical pharmacokinetics of cyclophosphamide. *Clin Pharmacokinet*, **4**: 380-394.
- Hashmi FK, Hussain K, Islam M, Ali M, Tipu MK, Khan MT and Latif A (2012). Cisplatin-induced nephrotoxicity in different regimens of cancer chemotherapy. *Health Med.*, **6**: 1917-1924
- Hauser E, Seidl R, Freilinger M, Male C and Herkner K (1996). Hematologic manifestations and impaired liver synthetic function during valproate monotherapy. *Brain Dev.* 18: 105-109.
- Hayden K and Van Heyningen C (2001). Measurement of total protein is a useful inclusion in liver function test profiles. *Clinical Chemistry*, **47**: 793-794.
- Holt MP and Ju C (2006). Mechanisms of drug-induced liver injury. *The AAPS Journal*, **8**: E48-E54.
- Injac R, Boskovic M, Perse M, Koprivec-Furlan E, Cerar A, Djordjevic A and Strukelj B (2008). Acute doxorubicin nephrotoxicity in rats with malignant neoplasm can be successfully treated with fullerenol C60 (OH) 24 via suppression of oxidative stress. *Pharmacol Rep*, **60**: 742.
- Jassem J, Pieńkowski T, Płuzańska A, Jelic S, Gorbunova V, Mrsic-Krmpotic Z, Berzins J, Nagykalnai T, Wigler N and Renard J (2001). Doxorubicin and paclitaxel versus fluorouracil, doxorubicin and cyclophosphamide as first-line therapy for women with metastatic breast

- cancer: Final results of a randomized phase III multicenter trial. *J. Clin. Oncol.*, **19**: 1707-1715.
- Je H (2011). Guyton and Hall textbook of medical physiology. WB Saunders Company, Saunders, London, pp.15-17.
- Kalender Y, Yel M and Kalender S (2005). Doxorubicin hepatotoxicity and hepatic free radical metabolism in rats: The effects of vitamin E and catechin. *Toxicology*, **209**: 39-45.
- Klasco R (2009). Drugdex® System (electronic version). Thomson Micromedex. Greenwood Village, Colorado, USA. Accessed via http://www. thomsonhc. com on,1: 06-09.
- Klotz U (2009). Pharmacokinetics and drug metabolism in the elderly. *Drug Metab. Rev*, **41**: 67-76.
- Landsberg L, Young J, Kasper D, Braunwald E, Fauci A, Hauser S, Longo D and Jameson L (2005). Harrison's principles of internal medicine. McGraw-Hill Professionals, pp.1735-1739.
- Leblond R, Degowin R and Brown D (2004). History taking and the medical record. DeGowin's Diagnostic Examination. 8th ed., McGraw-Hill Companies, Inc., New York,pp.19-20.
- Levels A (2000). Evaluation of abnormal liver-enzyme results in asymptomatic patients. *N. Engl. J. Med*, **342**(17): 1266-1271.
- Lis CG, Grutsch JF, Vashi PG and Lammersfeld CA (2003). Is serum albumin an independent predictor of survival in patients with breast cancer? *J. Parenter. Enteral. Nutr.*, 27: 10-15.
- Mangoni AA and Jackson SH (2004). Age□related changes in pharmacokinetics and pharmacodynamics: Basic principles and practical applications. *Br. J. Clin. Pharmacol.*, **57**: 6-14.
- Maurel M and Rosenbaum J (2012). Closing the gap on drug induced liver injury. *Hepatology*, **56**: 781-783.
- McPherson RA and Pincus MR (2011). Henry's clinical diagnosis and management by laboratory methods, Elsevier Health Sciences, pp.123-137
- Meyer RM, Gospodarowicz MK, Connors JM, Pearcey RG, Wells WA, Winter JN, Horning SJ, Dar AR, Shustik C and Stewart DA (2012). ABVD alone versus radiation-based therapy in limited-stage Hodgkin's lymphoma. *N. Engl. J. Med.*, **366**: 399-408.
- Miller LL, Bly C, Watson M and Bale W (1951). The dominant role of the liver in plasma protein synthesis a direct study of the isolated perfused rat liver with the aid of lysine-J-C14. *J. Exp. Med.*, **94**: 431-453.
- Murray K, Rodwell V, Bender D, Botham KM, Weil PA and Kennelly PJ (2009). Harper's Illustrated Biochemistry, McGraw-Hill, New York, p.28.
- Ong JP, Aggarwal A, Krieger D, Easley KA, Karafa MT, Van Lente F, Arroliga AC and Mullen KD (2003). Correlation between ammonia levels and the severity of hepatic encephalopathy. *Am. J. Med.*, **114**: 188-193.

- Pavek P and Dvorak Z (2008). Xenobiotic-induced transcriptional regulation of xenobiotic metabolizing enzymes of the cytochrome P450 super family in human extra hepatic tissues. *Curr. Drug Metab.*, **9**: 129-143.
- Pieniążek A, Czepas J, Piasecka-Zelga J, Gwoździński K and Koceva-Chyła A (2013). Oxidative stress induced in rat liver by anticancer drugs doxorubicin, paclitaxel and docetaxel. *Adv Med Sci*, 24-31.
- Pratt DS and Kaplan MM (2000). Evaluation of abnormal liver-enzyme results in asymptomatic patients. *N. Engl. J. Med.*, **342**: 1266-1271.
- Rashid S, Ali N, Nafees S, Ahmad ST, Arjumand W, Hasan SK and Sultana S (2013). Alleviation of doxorubicin-induced nephrotoxicity and hepatotoxicity by chrysin in Wistar rats. *Toxicol. Mech. Methods.*, 23: 337-345.
- Rolan P (1994). Plasma protein binding displacement interactions-why are they still regarded as clinically important? *Br. J. Clin. Pharmacol.*, **37**: 125.
- Sallie R, Michael Tredger J and Williams R (1991). Drugs and the liver Part 1: Testing liver function. *Biopharma*. *Drug Dispos.*, **12**: 251-259.
- Shargel L, Andrew B and Wu-Pong S (2005). Applied biopharmaceutics and pharmacokinetics, Appleton & Lange Reviews/McGraw-Hill, Medical Pub. Division.
- Shi S and Klotz U (2011). Age-related changes in pharmacokinetics. *Curr Drug Metab*, **12**: 601-610.
- Sokoll LJ and Dawson-Hughes B (1989). Effect of menopause and aging on serum total and ionized calcium and protein concentrations. *Calcified Tissue International.*, **44**: 181-185.
- Sørensen J, Klee M, Palshof T and Hansen H (1993). Performance status assessment in cancer patients. An inter-observer variability study. Br. J. Cancer. 67: 773.
- Steinherz LJ, Graham T, Hurwitz R, Sondheimer HM, Schwartz RG, Shaffer EM, Sandor G, Benson L and Williams R (1992). Guidelines for cardiac monitoring

- of children during and after anthracycline therapy: Report of the Cardiology Committee of the Childrens Cancer Study Group. *Pediatrics*, **89**: 942-949.
- Subramaniam SR, Cader RA, Mohd R, Yen KW and Ghafor HA (2013). Low-dose cyclophosphamide-induced acute hepatotoxicity. *Am. J. Case Rep.*, **14**: 345.
- Swerdlow AJ, Higgins CD, Smith P, Cunningham D, Hancock BW, Horwich A, Hoskin PJ, Lister A, Radford JA and Rohatiner AZ (2007). Myocardial infarction mortality risk after treatment for Hodgkin disease: A collaborative British cohort study. *J. Natl. Cancer Inst.*, **99**: 206-214.
- Throop JL, Kerl ME and Cohn LA (2004). Albumin in health and disease: Causes and treatment of hypoalbuminemia. *Compend. Contin. Educ. Pract. Vet.*, **26**: 940-949.
- Tranum B, Mcdonald B, Thigpen T, Vaughn C, Wilson H, Maloney T, Costanzi J, Bickers J, El Mawli NG & Palmer R (1982). Adriamycin combinations in advanced breast cancer: A southwest oncology group study. *Cancer*, **49**: 835-839.
- Triantafyllou K, Vlachogiannakos J and Ladas SD (2010). Gastrointestinal and liver side effects of drugs in elderly patients. *Best Pract. Res. Clin. Gastroenterol.*, **24**: 203-215.
- Turell L, Radi R and Alvarez B (2013). The thiol pool in human plasma: The central contribution of albumin to redox processes. *Free Radic. Biol. Med.*, **65**: 244-253.
- Turnheim K (2003). When drug therapy gets old: Pharmacokinetics and pharmacodynamics in the elderly. *Exp. Gerontol*, **38**: 843-853.
- Wolfe RR, Jahoor F and Hartl WH (1989). Protein and amino acid metabolism after injury. *Diabetes Metab. Rev.*, **5**: 149-164.
- Yoo KY (2010). Cancer prevention in the Asia pacific region. *Asian Pac. J. Cancer Prev.*, **11**: 839-844.