

Heterochelates of metals as an effective anti - Urease agents couple with their docking studies

Noshab Qamar^{1*}, Hira Sultan², Ahmad Raheel³, Maria Ashfaq¹, Rafia Azmat¹, Raheela Naz¹, Mehreen Lateef⁴, Khalid Mohammed Khan^{5,6} and Tanzila Arshad⁷

¹Department of Chemistry, University of Karachi, Karachi, Pakistan

²Department of Chemistry, NED University of Engineering and Technology, Karachi, Pakistan

³Department of Chemistry, Quaid-e-Azam University, Islamabad, Pakistan

⁴MDRL, Baharia University of Medical & Dental College, Karachi, Pakistan

⁵HEJ Research Institute of Chemistry, International Center of Chemical and Biological Sciences (ICCBS), University of Karachi, Karachi, Pakistan

⁶Department of Clinical Pharmacy, Institute for Research and Medical Consultations (IRMC), Imam Abdulrahman Bin Faisal University, Dammam, Saudi Arabia

⁷Department of Applied Chemistry and Chemical Technology, University of Karachi, Karachi, Pakistan

Abstract: The current article discusses the activities of several synthesized metal heterochelates in in-vitro as anti-ulcer agents followed by their docking study. For this purpose, two important ligands like 8-hydroxyquinoline and *DL*-methionine were used in synthesis of heterochelates of metal including Cr (III), Mn (II), Fe (III), Co (II), Ni (II), Cu (II), Zn (II), Cd (II) and Pb (II). It was observed that these complexes showed excellent urease inhibition activities in which thiourea was the standard having IC₅₀ value 21.6 ± 0.12 μM. The Cu (II) complex showed potent inhibitory activity (22.6 ± 0.72 μM) when compared with the standard thiourea (21.6 ± 0.12 μM) among the nine synthesized complexes while Mn (II), Fe (III), Cd (II) and Pb (II) also showed better inhibitory activities. The urease inhibitory activities of heterochelates also tested and validated by docking analysis.

Keywords: Urease, inhibition activity, complexes, docking.

INTRODUCTION

Urease is referred to as a Ni-containing metalloenzyme used to catalyze the hydrolysis of urea into ammonia (NH₃) and carbon dioxide (CO₂). It is naturally present in plant seeds (jack beans, soybeans, etc.), animal tissues and soil microorganisms. Hydrolysis of urea is a slow reaction while in the presence of urease enzyme, the rate is increased by 10¹⁴ times than nonenzymatic hydrolysis (Mobley *et al.*, 1995). Urease mechanism is responsible for the development of many diseases in humans like ulcer, urolithiasis, pyelonephritis, ammonia and hepatic encephalopathy, hepatic coma, and urinary catheter encrustation (Bremner *et al.*, 1989, Arshad *et al.*, 2018, Kajiwarra 2003). A Gram-negative bacteria, *Helicobacter pylori*, produces urease enzyme which causes peptic ulcer and stones by using urease mechanism (Bremner *et al.*, 1989). In soil, urease enzyme is produced by some microorganism (genus *Proteus*, *Klebsiella*, *Bacillus* and *Sporosarcina*) (Phang *et al.*, 2018); hydrolyzed product of urea is further converted into ammonium salt which is a basic nutrient for the plants. However, the excessive urease activity increases the production of ammonia which increases the ammonia toxicity in the environment as well as increases the consumption of urea (Sahrawat 1980, Bremner *et al.*, 1989). Urease inhibitors are used to reduce the activity of urease enzyme by decreasing the

hydrolysis of urea. This inhibition results in the decrement of *H. Pylori* survival inside the mucosa of the stomach. Discovery of new urease inhibitors is the active area of research to overcome all losses and health problems. A number of urease inhibitors have been reported which includes heavy metal ions (Habala *et al.*, 2018), hydroxamate derivatives (Font *et al.* 2008), phosphoramidites, urea derivatives, polyphenol (Zizian *et al.*, 2012), thiols (Benini *et al.* 1998), boric acid (Krajewska *et al.*, 1997), heterocyclic compounds (Karaali *et al.*, 2018), Schiff base (Sangeeta *et al.*, 2018), and phosphate (Krajewska and Ciurli., 2005). However, metal ion and their complexes with biologically active ligands are widely used to inhibit urease. Literature proved that heterochelates of many metals are known for biologically active nature for pathogenic microorganisms (Thakkar 2000) and play a significant part in the activation of enzymes. Usually, it has been seen that the complexes which have oxine ligand as a primary ligand are biologically active (Thakkar and Howard-Lock 1987). It was also reported that metal complexes of amino acids as ligands have significant biological and enzymatic properties (Perrin and Agarwal 1973). The aims and objectives of the current investigation are to search for a new horizon in the synthesis of metal complexes with their activities for controlling the significant diseases. For this purpose, synthesized complexes used in-vitro to monitor the urease activity. The chemical docking studies

*Corresponding author: e-mail: noshabqamar@gmail.com

of new complexes were also discussed in the relevant section of this article.

MATERIALS AND METHODS

Inhibition activity and docking of Heterochelates

All solvents and chemicals used as received. Heterochelates (1-9) synthesized by using Cr, Mn, Fe, Co, Ni, Cu, Zn, Cd and Pb metal with L₁ (8-Hydroxyquinoline) and L₂ (DL-Methionine). Synthesis and characterizations of several hetrochelates conducted by the method described by (Patel *et al.*, 2012) and their structure reported in the (fig. 1).

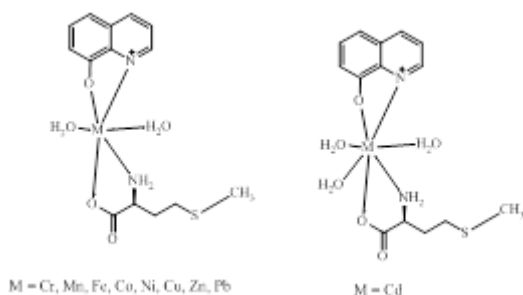


Fig. 1: The structure of synthesized metals complexes

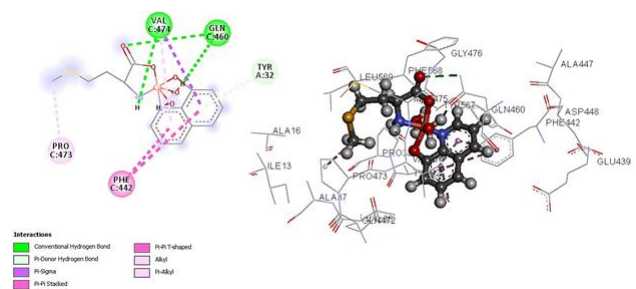


Fig. 2: 2D and 3D depiction of docking of (6) in active site of *Bacillus pasteurii* urease.

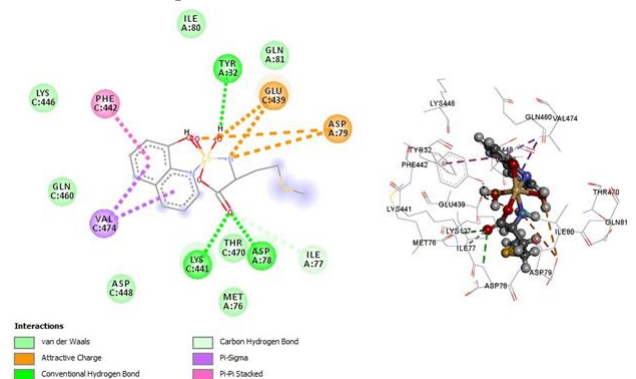


Fig. 3: 2D and 3D depiction of docking of (8) in active site of *Bacillus pasteurii* urease.

Urease inhibition activity

The urease inhibitory activity was resolved by a modified indophenol method described by (Lateef *et al.*, 2012) in triplicates which established quantity of ammonia produced during the reaction, results an increase in

absorbance after 50 min which was recorded at 630 nm on the micro plate reader. The thiourea was used as standard in this assay while inhibitions percentage determined through following formula:

$$\text{Inhibition (\%)} = \frac{(\text{OD control} - \text{OD test comp})}{\text{OD control}} \times 100$$

The IC₅₀ values were then calculated using the EZ-Fit Enzyme Kinetics program (Perrella Scientific Inc. Amherst, MA, USA).

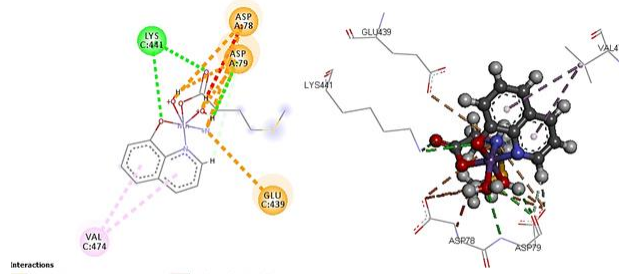


Fig. 4: 2D and 3D depiction of docking of (2) in active site of *Bacillus pasteurii* urease.

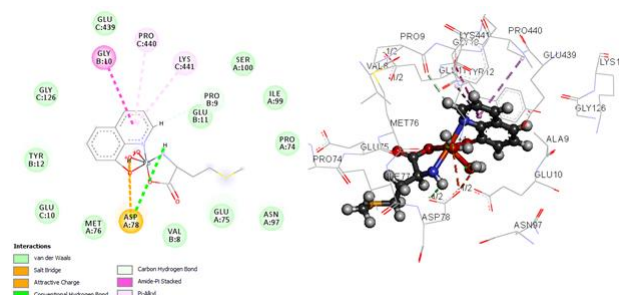


Fig. 5: 2D and 3D depiction of docking of (9) in active site of *Bacillus pasteurii* urease.

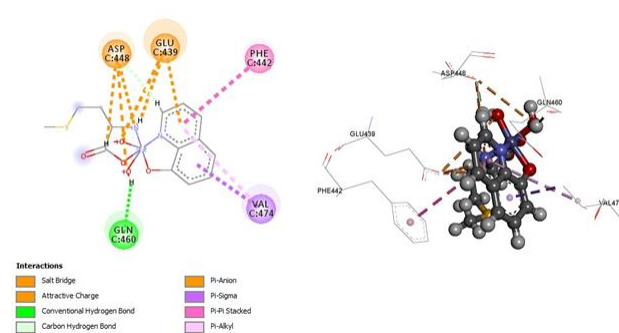


Fig. 6: 2D and 3D depiction of docking of (3) in active site of *Bacillus pasteurii* urease.

Molecular docking study

Autodock (Morris *et al.*, 2009) was used to perform the docking simulations employing the 3D structure of *Bacillus pasteurii* urease (PDB code: 4ubp). The Protein Data Bank was used to retrieve the “pdb” file (www.pdb.org). After the removal of all the het-atoms and the ligand, the Autodock tools were used to convert the protein to pdbqt format (1.5.6) (Morris *et al.*, 2009). For the preparation of ligand, the Marvin sketch (5.8.3)

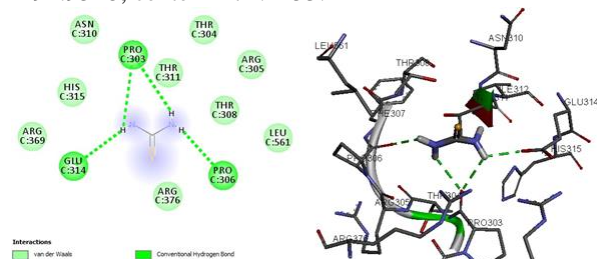
Table 1: Urease inhibition activity of Ligands and Compounds 1-9

Compound	Molecular formula	Mol. wt. (g/mol)	Urease Inhibition IC ₅₀ (μM)
L1			75.86 ± 0.82
L2			49.9 ± 0.20
1	C ₁₄ H ₂₀ CrN ₂ O ₅ S	380.05	> 200 μM
2	C ₁₄ H ₁₆ MnN ₂ O ₃ S	347.29	57.4 ± 0.44
3	C ₁₄ H ₁₆ FeN ₂ O ₃ S	348.20	69.0 ± 0.34
4	C ₁₄ H ₁₆ CoN ₂ O ₃ S	351.29	> 200 μM
5	C ₁₄ H ₁₆ NiN ₂ O ₃ S	351.05	> 200 μM
6	C ₁₄ H ₁₆ CuN ₂ O ₃ S	355.90	22.6 ± 0.72
7	C ₁₄ H ₁₆ ZnN ₂ O ₃ S	357.73	> 200 μM
8	C ₁₄ H ₂₁ CdN ₂ O ₆ S	458.81	53.7 ± 0.34
9	C ₁₄ H ₂₀ N ₂ O ₅ PbS	535.58	64.5 ± 0.82
Thiourea			21.6 ± 0.12

Table 2: Binding energies of synthesized compounds with bacillus pasteurii urease

Compound	Urease Inhibition IC ₅₀ (μM)	Binding affinity (kcal/mol)
2	57.4 ± 0.44	-7.0
3	69.0 ± 0.34	-6.7
6	22.6 ± 0.72	-8.2
8	53.7 ± 0.34	-7.2
9	64.5 ± 0.82	-9.7

was used for two-dimensional chemical structures of ligands (<http://www.chemaxon.com>) and these structures were then converted to 3D format by the Open Babel (ver 2.3.1) (O'Boyle *et al.*, 2011). Finally, Autodock Tools were used to prepare the final pdbqt format of ligands. The following parameters were used to perform the docking simulation: size x=82.6443; size y=86.0453.2962; size z=67.2583; center x=15.5310; center y=94.9548; center z=74.2453.

**Fig. 7:** 2D and 3D depiction of docking of in active site of *Bacillus pasteurii* urease.

RESULTS

The molecular formula of all synthesized complexes (1-9) using two ligands were reported in the table 1 which were subjected to urease activity followed by in-silico or docking studies, while their results are expressed in the form of IC₅₀ values (table 1). The IC₅₀ values of both ligands compared with a standard Thiourea (IC₅₀ = 21.6 ± 0.12), displayed urease inhibition activity. The superior activities of synthesized complexes were observed when compared with the standard Thiourea, where compounds 2, 3, 8 and 9 were found to be active except compounds 1

and 7 (IC₅₀ > 200 μM). While compound 6 containing Cu metal (IC₅₀ = 22.6 ± 0.72) exhibited significant inhibitory potential.

The interaction patterns of the synthesized compounds with urease enzyme were investigated through molecular docking studies. The Auto Dock tool was employed in order to calculate the preminent conformational place of the synthesized compounds against *Bacillus pasteurii* urease (PDB code: 4ubp). Only those compounds were analyzed, which were already active against urease in-vitro activity. The analysis of the generated docked complexes was done by the minimum energy values (kcal/mol) and bonding interaction pattern (hydrogen/hydrophobic). Docking results revealed that the compound (6) showed the lowest energy value (-8.2 kcal/mol). Docked complex of compound (3) has predicted -6.7 kcal/mol energy value, which was comparatively higher than other synthesized compounds as shown in Table 2.

DISCUSSION

According to the result of urease inhibition activity, the compound 6 containing copper (II) ion when coordinated with the active site of nickel-dependent enzyme urease, fallouts in the alteration of the active site (Qiu *et al.* 2013, Sun *et al.* 2012, Mumtaz *et al.* 2018). The strong inhibitory effect related to ureolytic activity through synthesized copper complex may be explained as i) might be due to the strong Lewis acid properties of Copper (II) ions (Li Y-G *et al.*, 2007), ii) its binding to histidine residues in proteins, iii) polymerization of

protein iv), and blocking of the thiol group (Follmer *et al.*, 2005). It has also been observed that compounds containing O and N donor site displayed greater interaction and formation of H-bonding with the urease enzyme (Rauf *et al.*, 2011, Ghous *et al.*, 2010, Akhtar *et al.*, 2011).

The following interaction through analysis of docked complexes were observed, which were hydrogen bonding, Pi-Alkyl, Pi-Sulfur, Pi-anion, Pi-sigma, and Van der Waals. Docking analysis showed that all the docked compounds interacted with active binding region of *Bacillus pasteurii* urease (Kafarski *et al.*, 2018 29). Different interactions in docked complexes and the 2D graphical depictions are presented in Fig. (2-6). Docking results of the standard drug (thiourea) showed that it was strongly bound to the active site of the urease enzyme and formed three conventional hydrogen bonds with PRO C:303, PRO C:306 and GLU C:314. Van-der-Waal interactions were also shown by thiourea with the urease structure. These interactions bind thiourea with the *Bacillus pasteurii* urease, as a result thiourea acts as a good inhibitor of urease (fig. 7).

Higher anti-urease activity of (6) against urease obtained from experimental results is also elucidated from docking results as (6) strongly binds to *Bacillus pasteurii* urease and form multiple conventional hydrogen bonds with GLN C:460, and VAL C:474 along with other Pi-Pi, Pi-alkyl and alkyl interactions with different other amino acid groups (as shown in Fig. 2). Docking results showed that the binding energy value of (6) is lower than all other compounds and this data is in good agreement with the experimental data. In case of (8), it showed relatively higher binding energy (-7.2 kcal/mol) than (6) and formed hydrogen bonds with TYR A: 32 and LSS C441 along with some attractive interactions with other amino acid residues of the urease enzyme (Fig. 3). Similar interaction patterns were also shown by compounds (2, 3 and 9).

CONCLUSION

The inhibitory activity of metal compounds has been a subject of interest in the current study. The IC₅₀ of all hetero-chelates were compared with the standard. Moreover, it was found that the copper compound exhibited significant urease inhibition activities. The docking study of these heterochelates also showed that the compound containing copper as its center metal found to have greater potential in the inhibitory the effect of urease enzyme as compare to all synthesized hetero-chelates. Due to its highest activity it is recommended as future potent anti-ulcer agents.

REFERENCES

Akhtar M, Iqbal L, Lateef M, Nawab B, Saleem M and Afza N (2011). Bio-Reactive Properties of Citrus

- Waste: An Investigation of Antioxidant and Tyrosinase Inhibitory Activities. *Pak. J. Bot.*, **43**(6): 2881-2883.
- Arshad T, Sheikh HK, Kazmi MH, Farheen S, Sohail T and Lateef M (2018). New bioactive triaryl triglyceride esters: Synthesis, characterization and biological activities. *Bangladesh J. Pharmacol.*, **13**(4): 302-308.
- Benini S, Rypniewski W, Wilson K, Ciurli S, Mangani S (1998). The complex of *Bacillus pasteurii* urease with β -mercaptoethanol from X-ray data at 1.65-Å resolution. *J. Biol. Inorg. Chem.*, **3**: 268-273.
- Bremner JM and Krogmeier MJ (1989). Evidence that the adverse effect of urea fertilizer on seed germination in soil is due to ammonia formed through hydrolysis of urea by soil urease. *Proc. Natl. Acad. Sci., USA.* **86**: 8185-8188.
- Follmer C and Carlini CR (2005). Effect of chemical modification of histidines on the copper-induced oligomerization of jack bean urease (EC 3.5.1.5). *Arch. Biochem. Biophys.*, **435**(1): 15-20.
- Font M, Domínguez, M a.-J, Sanmartín C, Palop JA, San-Francisco S, Urrutia O, Houdusse F, García-Mina JM (2008). Structural characteristics of phosphoramidate derivatives as urease inhibitors. Requirements for activity. *J. Agric. Food Chem.*, **56**: 8451-8460.
- Ghous T, Akhtar K, Nasim F and Choudhry MA (2010). Screening of selected medicinal plants for urease inhibitory activity. *Biol. Med.*, **2**(4): 64-69.
- Habala L, Devínsky F and Egger AE (2018). REVIEW: Metal complexes as urease inhibitors. *J. Coord. Chem.*, **71**(7): 907-940.
- Howard-Lock HE (1987). Lock CJL in *Comprehensive Co-ordination Chemistry*, Wilkinson G, Gillard RD, McCleverty JA, Eds. Pergamon Press, Oxford. **6**: 755.
- Kafarski, Paweł, and Talma M (2018). Recent advances in design of new urease inhibitors: A review. *J. Adv. Res.*, **13**: 101-112.
- Kajiwarra M (2003). U.S. Patent Application No. 10/669, 700.
- Karaali N, Menteşe M, Baltas N and Mentese E (2018). Synthesis and urease inhibition study of some new quinazolinone derivatives bearing triazole, thiadiazole, and piperazine moiety. *J. Heterocycl. Chem.*, **55**(11): 2571-2577.
- Krajewska B and Ciurli S (2005). Jack bean (*Canavalia ensiformis*) urease. Probing acid-base groups of the active site by pH variation. *Plant Physiol. Biochem.*, **43**: 651-658.
- Krajewska B, Zaborska W and Leszko M (1997). Inhibition of chitosan-immobilized urease by boric acid as determined by integration methods. *J. Mol. Catal. B: Enzym.*, **3**: 231-238.
- Lateef M, Iqbal L, Fatima N, Siddiqui K, Afza N, Zia-ul-Haq M and Ahmed M (2012). Evaluation of antioxidant and urease inhibition activities of roots of *Glycyrrhiza glabra*. *Pak. J. Pharm. Sci.*, **25**(1): 99-102.
- Li YG, Shi DH, Zhu HL, Yan H and Ng SW (2007). Transition metal complexes (M = Cu, Ni and Mn) of

- Schiff-base ligands: Syntheses, crystal structures and inhibitory bioactivities against urease and xanthine oxidase. *Inorg Chim Acta*, **360**(9): 2881-2889.
- Mobley H, Island MD, Hausinger RP (1995). Molecular biology of microbial ureases. *Microbiol. Rev.*, **59**: 451-480.
- Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS and Olson AJ (2009). AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. comput. Chem.*, **30**: 2785-2791.
- Mumtaz A, Arshad J, Saeed A, Nawaz MAH and Iqbal J (2018). Synthesis, Characterization and Urease Inhibition Studies Of Transition Metal Complexes Of Thioureas Bearing Ibuprofen Moiety, *J. Chil. Chem. Soc.*, **63**(2): 3934-3940.
- O'Boyle NM, Banck M, James CA, Morley C, Vandermeersch T and Hutchison GR (2011). Open Babel: An open chemical toolbox. *J. Cheminform.*, **3**: 33.
- Patel AD, Prajapati NK and Vora JJ (2012). Synthesis and characterization of transition metal heterochelates with oxine and vitamin U. *Der Pharmacia Sinica.*, **3**(1): 93-98.
- Perrin DD and Agarwal RP (1973). Metal ions in biological systems. Ed. Sigel HC, Marcel Dekker, New York. **2**: 167.
- Phang IR, Chan YS, Wong KS, Lau SY, (2018). Isolation and characterization of urease-producing bacteria from tropical peat. *Biocatal. Agric. Biotechnol.*, **13**: 168-175.
- Qiu X, Wang J, Shi D, Li S, Zhang F, Zhang F, Cao G and Zhai B (2013). Syntheses, urease inhibition activities and fluorescent properties of transition metal complexes. *J. Coord. Chem.*, **66**(9): 1616-1625.
- Rauf A, Ahmed F, Qureshi AM, Aziz-ur-Rehman, Khan A, Qadir MI, Choudary MI, Chohan ZH, Youssoufif MH and Haddad TB (2011). Synthesis and Anti-Urease Activity of Novel Thio/Barbiturates. *J. Chin. Chem. Soc.*, **58**: 528-537.
- Sahrawat KL (1980). Control of urea hydrolysis and nitrification in soil by chemicals. Prospects and problems. *Plant Soil*. **57**: 335-352.
- Sangeeta S, Ahmad K, Noorussabah N, Bharti S, Mishra M, Sharma S and Choudhary M (2018). Synthesis, crystal structures, molecular docking and urease inhibition studies of Ni(II) and Cu(II) Schiff base complexes. *J. Mol. Struct.* **1156**: 1-11.
- Sun L, Li Y, Dong X and Guo M (2012). Synthesis, characterization, and urease inhibitory activity of two copper(II) complexes of cyclohexanecarboxylate. *Transit. Metal Chem.*, **37**: 361-366.
- Thakkar JR and Thakkar NV (2000). Synthesis and characterization of chiral, mixed Ligand Co(II) complexes of isonitrosopropiophenone and amino acids. *Syn. React. Inorg. Metal-Org. Chem.*, **30**: 1871-1887.
- Zizian H, Nabati F, Sharifi A, Siavoshi F, Mahdavi M and Amanlou M (2012). Large-scale virtual screening for the identification of new helicobacter pylori urease inhibitor scaffolds. *J. Mol. Model.*, **18**: 2917-2927.