

***N*-Sulfonated derivatives of (2-furoyl)piperazine: Promising antibacterial agents with mild cytotoxicity**

Muhammad Athar Abbasi^{1,*}, Misbah Irshad², Aziz-ur-Rehman¹, Sabahat Zahra Siddiqui¹, Syed Adnan Ali Shah³ and Muhammad Shahid⁴

¹Department of Chemistry, Government College University, Lahore, Pakistan

²Department of Chemistry, Division of Science and Technology, University of Education, Township Lahore, Pakistan

³Faculty of Pharmacy and Atta-ur-Rahman Institute for Natural Products Discovery (AuRIns), Level 9, FF3, Universiti Teknologi MARA, Puncak Alam Campus, 42300 Bandar Puncak Alam, Selangor Darul Ehsan, Malaysia

⁴Department of Biochemistry, University of Agriculture, Faisalabad, Pakistan

Abstract: 2-Furoyl-1-piperazine (1) was treated with a series of alkyl/aryl sulfonyl chlorides (2a-i) under benignant conditions to obtain its *N*-sulfonated derivatives (3a-i). These compounds were screened for their antibacterial potential against pathogenic bacteria. The low Minimum Inhibitory Concentration (MIC) values of these molecules, in comparison of ciprofloxacin, demonstrated their high antibacterial potential. Cytotoxic activities were ascertained through their hemolytic potential and mild hemolytic profiles of these compounds proved them to be promising compounds for drug designing and development.

Keywords: Piperazine, furan, sulfonamides, antibacterial, hemolytic activity.

INTRODUCTION

The discovery of new drug molecules through natural or synthetic routes is ongoing research from a number of years. Similarly, some biologically active compounds have been modified to synthesize new compounds consisting of active pharmacophores (Mohs and Greig, 2017). Organic chemists have employed piperazine (1,4-diazinane) to synthesize different derivatives with more bioactive potential. The applications of piperazine in medicinal chemistry include the synthesis of antimicrobial (Jalager *et al.*, 2019; Tang *et al.*, 2020) and anti-viral agents (Dou *et al.*, 2012). The piperazine derivatives also have vast applications in the fields of engineering (Bruder *et al.*, 2011) and polymer (Yamaguchi *et al.*, 2010).

Sulfonamides bear sulfonyl group (-SO₂N<) (Ordones *et al.*, 2011). Sulfonamides are good antibacterial agents because the sulfonyl functionality perturbs folic acid synthesis and thus reduces the bacterial growth (El-Sayed *et al.*, 2011; Ozbek *et al.*, 2019; Saeedi *et al.*, 2014). Sulfonamide compounds have been used as preventive agents in chemotherapy against various diseases. These are also reported to possess antifungal, anti-inflammatory, antioxidant and enzyme inhibitory activities (Thach *et al.*, 2021; Abbasi *et al.*, 2020^a; El-Sayed *et al.*, 2011; Hassan *et al.*, 2021; Zoumpoulakis *et al.*, 2012). Moreover, these are broadly used as anticancer (Mun *et al.*, 2012), antiviral agents as well as effective inhibitor against HIV protease (Clercq *et al.*, 2001). Sulfonamides have been found as biologically active as they have shown significant cytotoxic effects against breast cancer cells

(Mirian *et al.*, 2011). A list of infections including livestock herbs, urinary tract and gastrointestinal ones was also treated by sulfonamides (Root *et al.*, 2021; Apaydin *et al.*, 2019). So, the immense therapeutic potential of sulfonamides prompted us to synthesize some new molecules bearing piperazine functionality and to explore their antibacterial potentials and hemolytic behaviors.

MATERIALS AND METHODS

All chemicals and reagents required for experimental work were bought from Alfa Aesar and sigma Aldrich. Analytical grade solvents were provided by local suppliers. Griffin and George apparatus were used to collect melting point data by using open capillary method. Thin layer chromatography plates were utilized for determination of initial purity of compounds at 254 nm. Gradient solvent system used was comprised of ethyl acetate and n-hexane in 30:70 and played a role as mobile phase. IR spectra were recorded by KBr pellet method on a Jasco-320-A spectrometer. Bruker spectrometers were used for recording ¹H-NMR signals at 400 and 600 MHz. CDCl₃ was solvent used for this purpose. For the purpose of recording ¹³C-NMR signals at 150 MHz in CDCl₃ Bruker spectrometer was used.

Synthesis of 4-(alkyl/aryl sulfonyl)-1-(2-furoyl)piperazine (3a-i)

Suspension of 2-furoyl-I-piperazine (22.0 mmol; 1) was prepared by dispersing it in 20.0 mL of distilled water, taken in a 100 ml round bottom flask. 10% Na₂CO₃ solution was used to adjust the pH of suspension towards alkalinity in range 9-10. Then different alkyl/aryl sulfonyl chlorides (22.0 mmol; 2a-i) were gradually added in small patches, one in each reaction. Reaction mixture was

*Corresponding author: e-mail: abbasi@gcu.edu.pk

stirred for 3 to 4 hours. Progress of the reaction was monitored by TLC. Mixture was allowed to stand for 60 minutes after adding ice cold distilled water in excess, thereby, precipitates were formed in good yield. After filtration, the obtained precipitates were washed with distilled water and then air dried to acquire title compounds in purified form.

Physical and Spectral Characterization

4-(Methylsulfonyl)-1-(2-furoyl)piperazine (3a)

White amorphous solid; Yield: 87%; m.p.: 94-96°C; Mol. For.: C₁₀H₁₄N₂O₄S; Mol. Mass.: 258 g/mol; IR (KBr, ν , cm⁻¹): 3027 (Ar C-H), 2836 (R C-H), 1603 (C=O), 1584, 1453 (C=C), 1323 (S=O), 1156 (C-O-C), 1078 (C-N-C), 655 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 7.43 (dist. d, J = 0.9 Hz, 1H, H-5), 7.02 (d, J = 2.6 Hz, 1H, H-3), 6.45 (dd, J = 1.8, 3.4 Hz, 1H, H-4), 3.87 (br.s, 4H, CH₂-3' & CH₂-5'), 3.24 (br.t, J = 5.1 Hz, 4H, CH₂-2' & CH₂-6'), 2.73 (s, 3H, CH₃-1"); ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): 159.1 (C-6), 147.4 (C-2), 144.0 (C-5), 117.5 (C-4), 111.6 (C-3), 45.9 (C-2', C-3', C-5', C-6'), 34.7 (C-1", merged owing to ring flipping).

4-(2-Naphthylsulfonyl)-1-(2-furoyl)piperazine (3b)

White amorphous solid; Yield: 90%; m.p.: 91-93°C; Mol. For.: C₁₉H₁₈N₂O₄S; Mol. Mass.: 370 g/mol; IR (KBr, ν , cm⁻¹): 3038 (Ar C-H), 2835 (R C-H), 1612 (C=O), 1587 (C=C), 1333 (S=O), 1198 (C-O-C), 1161 (C-N-C), 657 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 8.35 (br.s, 1H, H-1"), 8.01-7.98 (m, 1H, H-8"), 7.95 (br.d, J = 8.1 Hz, 1H, H-5"), 7.70-7.64 (m, 4H, H-3", H-4", H-6", H-7"), 7.44 (dist. d, J = 1.6 Hz, 1H, H-5), 6.99 (d, J = 3.4 Hz, 1H, H-3), 6.46 (dd, J = 1.8, 3.4 Hz, 1H, H-4), 3.92 (br.s, 4H, CH₂-3' & CH₂-5'), 3.18 (dist. t, J = 5.1 Hz, 4H, CH₂-2' & CH₂-6'); ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): 158.8 (C-6), 147.4 (C-2), 143.8 (C-5), 135.0 (C-6"), 132.4 (C-1", C-9", merged), 132.2 (C-4"), 129.4 (C-5"), 129.2 (C-3", C-7"), 129.1 (C-10"), 127.9 (C-8"), 127.7 (C-2"), 117.3 (C-4), 111.4 (C-3), 46.2 (C-2', C-3', C-5', C-6', merged owing to ring flipping).

4-(Phenylsulfonyl)-1-(2-furoyl)piperazine (3c)

White amorphous solid; Yield: 85%; m.p.: 96-98°C; Mol. For.: C₁₅H₁₆N₂O₄S; Mol. Mass.: 320 g/mol; IR (KBr, ν , cm⁻¹): 3039 (Ar C-H), 2841 (R C-H), 1618 (C=O), 1588 (C=C), 1337 (S=O), 1159 (C-O-C), 1088 (C-N-C), 658 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 7.78 (d, J = 7.8 Hz, 2H, H-2", H-6"), 7.66 (t, J = 7.5 Hz, 1H, H-4"), 7.59 (t, J = 7.6 Hz, 2H, H-3", H-5"), 7.47 (br.s, 1H, H-5), 7.03 (d, J = 3.4 Hz, 1H, H-4), 6.49 (dd, J = 3.7, 2.1 Hz, 1H, H-3), 3.92 (br.s, 4H, CH₂-3', CH₂-5'), 3.12 (t, J = 5.0 Hz, 4H, CH₂-2', CH₂-6'); ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): 158.9 (C-6), 147.4 (C-2), 143.9 (C-5), 135.3 (C-1"), 133.2 (C-4"), 129.2 (C-3", C-5"), 127.7 (C-2", C-6"), 117.7 (C-4), 111.5 (C-3), 46.2 (C-2', C-3', C-5', C-6', merged owing to ring flipping).

4-(4-Acetamidophenylsulfonyl)-1-(2-furoyl)piperazine (3d)

White amorphous solid; Yield: 82%; m.p.: 92-94°C; Mol. For.: C₁₇H₁₉N₃O₅S; Mol. Mass.: 377 g/mol; IR (KBr, ν , cm⁻¹): 3038 (Ar C-H), 2839 (R C-H), 1652, 1606 (C=O), 1590 (C=C), 1389 (S=O), 1163 (C-O-C), 1175 (C-N-C), 656 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 7.93 (br.s, 1H, N-H), 7.70 (br.d, J =8.6 Hz, 2H, H-3", H-5"), 7.66 (t, J =6.5 Hz, 2H, H-2", H-6"), 7.45 (br. d, J =0.9 Hz, 1H, H-5), 6.99 (br. d, J =2.5 Hz, 1H, H-3), 6.46 (dd, J =3.4, 1.7 Hz, 1H, H-4), 3.88 (br.s, 4H, CH₂-3', CH₂-5'), 3.06 (t, J =5.0 Hz, 4H, CH₂-2', CH₂-6'), 2.19 (s, 3H, CH₃-8"); ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): 168.8 (C-7"), 158.9 (C-6), 147.2 (C-2), 144.0 (C-4"), 142.6 (C-5), 129.6 (C-1"), 128.9 (C-2", C-6"), 119.3 (C-3", C-5"), 117.4 (C-4), 111.5 (C-3), 46.2 (C-2', C-3', C-5', C-6', merged owing to ring flipping), 24.6 (C-8").

4-(4-Methylphenylsulfonyl)-1-(2-furoyl)piperazine (3e)

White amorphous solid; Yield: 80%; m.p.: 90-92°C; Mol. For.: C₁₆H₁₈N₂O₄S; Mol. Mass.: 334 g/mol; IR (KBr, ν , cm⁻¹): 3046 (Ar C-H), 2838 (R C-H), 1658 (C=O), 1581 (C=C), 1335 (S=O), 1157 (C-O-C), 1092 (C-N-C), 661 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 7.62 (d, J =6.4 Hz, 2H, H-2", H-6"), 7.44 (br. d, J = 0.9 Hz, 1H, H-5), 7.33 (d, J = 8.0 Hz, 2H, H-3", H-5"), 6.99 (dist.d, J = 2.6 Hz, 1H, H-3), 6.45 (dd, J = 3.4, 1.8 Hz, 1H, H-4), 3.88 (br.s, 4H, CH₂-3', CH₂-5'), 3.06 (t, J = 5.1 Hz, 4H, CH₂-2', CH₂-6'), 2.42 (s, 3H, CH₃-7"); ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): 158.8 (C-6), 147.4 (C-2), 144.1 (C-5), 143.9 (C-1"), 132.2 (C-4"), 129.8 (C-3", C-5"), 127.7 (C-2", C-6"), 117.3 (C-4), 111.5 (C-3), 46.2 (C-2', C-3', C-5', C-6', merged owing to ring flipping), 21.5 (C-7").

4-(4-Bromophenylsulfonyl)-1-(2-furoyl)piperazine (3f)

White amorphous solid; Yield: 88%; m.p.: 97-99°C; Mol. For.: C₁₅H₁₅BrN₂O₄S; Mol. Mass.: 398 g/mol; IR (KBr, ν , cm⁻¹): 3038 (Ar C-H), 2838 (R C-H), 1614 (C=O), 1587 (C=C), 1395 (S=O), 1162 (C-O-C), 1084 (C-N-C), 653 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 7.72 (d, J =6.6 Hz, 2H, H-2", H-6"), 7.64 (d, J =6.6 Hz, 2H, H-3", H-5"), 7.48 (dist.d, J =0.9 Hz, 1H, H-5), 7.05 (dist.d, J = 2.5 Hz, 1H, H-3), 6.50 (dd, J =3.4, 1.7 Hz, 1H, H-4), 3.93 (br. s, 4H, CH₂-3', CH₂-5'), 3.11 (t, J 5.1 Hz, 4H, CH₂-2', CH₂-6'); ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): ¹³C-NMR (150 MHz, CDCl₃, δ , ppm): 158.8 (C-6), 147.3 (C-2), 143.9 (C-5), 134.4 (C-1"), 132.4 (C-3", C-5"), 129.1 (C-2", C-6"), 128.4 (C-4"), 117.5 (C-4), 111.5 (C-3), 46.1 (C-2', C-3', C-5', C-6', merged owing to ring flipping).

4-(4-Acetylphenylsulfonyl)-1-(2-furoyl)piperazine (3g)

White amorphous solid; Yield: 86%; m.p.: 88-90 °C; Mol. For.: C₁₇H₁₈N₂O₅S; Mol. Mass: 362 g/mol; IR (KBr, ν , cm⁻¹): 3035 (Ar C-H), 2839 (R C-H), 1709, 1618 (C=O), 1587 (C=C), 1331 (S=O), 1159 (C-O-C), 1095 (C-N-C), 658 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ , ppm): 7.88 (d, J = 7.1 Hz, 2H, H-2", H-6"), 7.82 (d, J = 6.6 Hz, 2H, H-

3", H-5"), 7.49-7.47 (m, 1H, H-5), 7.05-7.03 (m, 1H, H-3), 6.50-6.48 (m, 1H, H-4), 3.94 (br. s, 4H, CH₂-3', CH₂-5'), 3.14 (t, *J* = 5.0 Hz, 4H, CH₂-2', CH₂-6'), 2.65 (s, 3H, CH₃-8"); ¹³C-NMR (150 MHz, CDCl₃, δ, ppm): 189.7 (C-7"), 158.9 (C-6), 147.3 (C-2), 144.0 (C-5), 142.5 (C-1"), 138.9 (C-4"), 135.8 (C-3", C-5"), 128.9 (C-2", C-6"), 117.5 (C-4), 111.6 (C-3), 46.2 (C-2', C-3', C-5', C-6', merged owing to ring flipping), 19.0 (C-8").

4-(2,4,6-Trimethylphenylsulfonyl)-1-(2-furoyl) piperazine (3h)

White amorphous solid; Yield: 84%; m.p.: 94-96°C; Mol. For.: C₁₈H₂₂N₂O₄S; Mol. Mass: 362 g/mol; IR (KBr, ν, cm⁻¹): 3088 (Ar C-H), 2846 (R C-H), 1619 (C=O), 1589 (C=C), 1329 (S=O), 1159 (C-O-C), 1081 (C-N-C), 657 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ, ppm): 7.49 (dist.d, *J*=0.9 Hz, 1H, H-5), 7.06 (dist.d, *J*=3.4 Hz, 1H, H-3), 6.99 (s, 2H, H-3" & H-5"), 6.51 (dd, *J*=1.8, 3.4 Hz, 1H, H-4), 3.86 (br.s, 4H, CH₂-3' & CH₂-5'), 3.27 (br.t, *J* 5.2 Hz, 4H, CH₂-2' & CH₂-6'), 2.65 (s, 6H, CH₃-7" & CH₃-9"), 2.65 (s, 3H, CH₃-8"); ¹³C-NMR (150 MHz, CDCl₃, δ, ppm): 159.0 (C-6), 147.5 (C-2), 143.9 (C-1"), 143.0 (C-5), 140.5 (C-2" & C-6"), 132.1 (C-4"), 130.9 (C-3" & C-5"), 117.2 (C-4), 111.5 (C-3), 44.5 (C-2', C-3', C-5', C-6', merged owing to ring flipping), 22.9 (C-7", C-8", C-9", merged).

4-(4-Methoxyphenylsulfonyl)-1-(2-furoyl)piperazine (3i)

White amorphous solid; Yield: 88%; m.p.: 90-92°C; Mol. For.: C₁₆H₁₈N₂O₅S; Mol. Mass: 350 g/mol; IR (KBr, ν, cm⁻¹): 3029 (Ar C-H), 2838 (R C-H), 1617 (C=O), 1581 (C=C), 1328 (S=O), 1159 (C-O-C), 1098 (C-N-C), 663 (C-S); ¹H-NMR (600 MHz, CDCl₃, δ, ppm): 7.71-7.69 (m, 4H, H-2", H-3", H-5", H-6"), 7.47 (dist.d, *J*=0.9 Hz, 1H, H-5), 7.03-7.01 (m, 1H, H-3), 6.49 (dd, *J*=3.4, 1.8 Hz, 1H, H-4), 3.91 (br.s, 4H, CH₂-3', CH₂-5'), 3.89 (m, 3H, CH₃-7"), 3.08 (t, *J*=5.0 Hz, 4H, CH₂-2', CH₂-6'); ¹³C-NMR (150 MHz, CDCl₃, δ, ppm): 163.3 (C-4"), 158.9 (C-6), 147.4 (C-2), 143.9 (C-5), 129.8 (C-1"), 126.7 (C-2", C-6"), 117.3 (C-4), 114.4 (C-3", C-5"), 111.5 (C-3), 46.2 (C-2', C-3', C-5', C-6' merged owing to ring flipping), 55.6 (C-7").

Antibacterial activity assay

Monitoring of synthesized compounds 3a-i was done for checking antibacterial activity against pathogenic bacteria. 96-Wells sterilized microplates were utilized for evaluating antimicrobial activity assay (Eloff et al., 1998). Test samples were dissolved in suitable solvent and taken in wells. Different samples of Tryptic Soy Broth (TSB) were taken in order to determine Minimum Inhibitory Concentration (MIC) values. The basic principle of determination corresponds to increase or decrease in total number of cells in life cycle of microbial growth. Optical density of samples was determined using 630 nm microplate reader Results were evaluated in terms of increase or decrease of absorption capacity of the tested medium (Kaspady et al., 2009; Yang et al., 2006).

Minimum inhibitory concentrations (MIC)

100μL of nutrient broth was poured in all wells of 96 well plates (micro dilution plates). Subsequently added 100μL of samples in the first well and used its two-fold diluted form by making the use of dilution method. Later on all the wells were cultured with 20μL of bacterial strains and incubation at 37°C for 24 hours were carried out. Resazurine was used as bacterial growth indicator and color changes from blue to pink scored as bacterial growth initiation (Abbasi et al., 2020^b).

STATISTICAL ANALYSIS

After three-folds performance and analysis of statistical data by using Microsoft Excel 2010, results are mentioned in terms of ± SEM. EZ Fit Perrella Scientific Inc. Amherst USA software was used to find out minimum inhibitory concentration (MIC) value under different dilutions values (ranging 5-30μg/well).

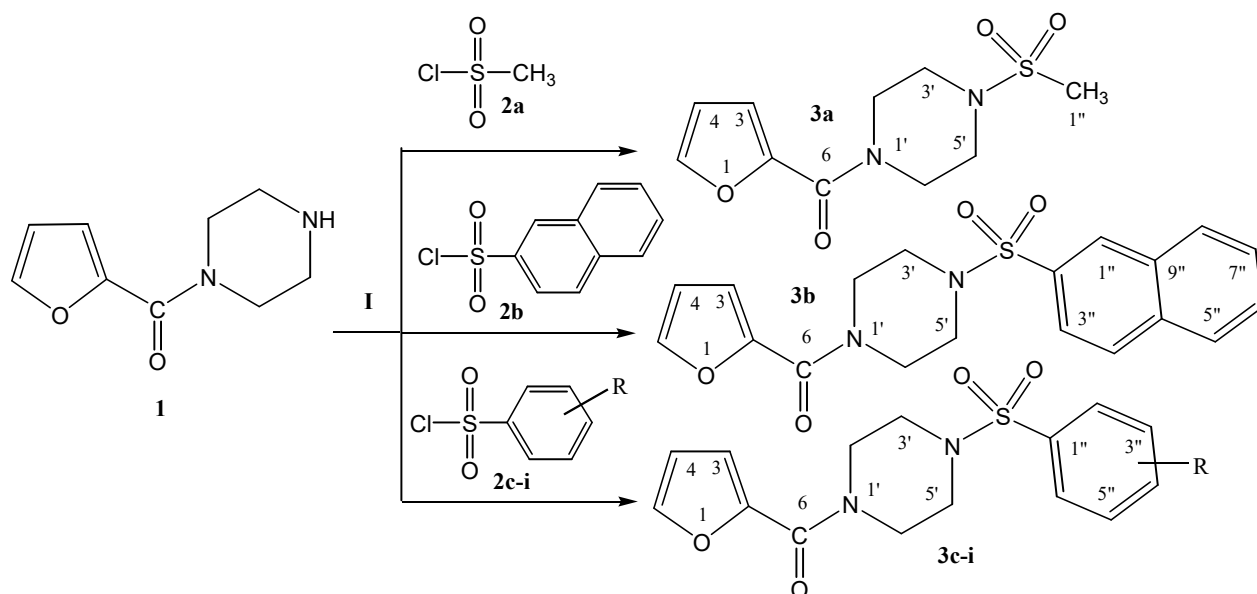
Hemolytic activity assay

The compounds were subjected to hemolytic activity analysis using reported method (Powell et al., 2000; Sharma et al., 2001; Rodríguez et al., 2017). Collection of fresh heparinized bovine blood was done from volunteers after approval and recommendation from Department of Clinical, Medicine and Surgery, University of Agriculture, Faisalabad, Pakistan. Samples in polystyrene tube were centrifuged at 1000xg for 10 min followed by removal of supernatant. Erythrocytes thus separated were washed and diluted with phosphate buffer to adjust the pH at 7.4. To this prepared suspension, synthetic compounds were added and cell count was determined by haemocytometer. Synthetic compounds (20μL of solution; one in each experiment) were added (made by 10mg/mL) in 180μL of Red Blood Cells (RBCs) suspension and incubated for 30 min at room temperature. Phosphate Buffered Saline (PBS) and Triton 100-X were used as negative and positive controls respectively (Shahid et al., 2013). The %age of hemolysis was taken as by using formula:

$$\% \text{ of Hemolysis} = \frac{\text{Absorbance of Sample} - \text{Absorbance of Negative Control}}{\text{Absorbance of Positive Control}} \times 100$$

RESULTS

The synthesis of targeted sulfonamides, 3a-i, is outlined in scheme 1. The method of synthesis is explicated in the experimental section. After structural corroboration, the compounds were subjected to antibacterial and hemolytic activities. The results of antibacterial activity of synthesized compounds are depicted as % inhibition values and MIC Minimum Inhibitory Concentration (table 1 & table 2). The hemolytic activity is given as % lysis in table 3.



Scheme 1: Outline for the synthesis of different sulfonamides (3a-i). Reagents & Conditions: (I) Aq. Na₂CO₃ soln./pH 9-10/stirring at room temperature (RT) for 3-4 hours.

Table 1: Antibacterial activity (inhibition %) of the synthesized compounds.

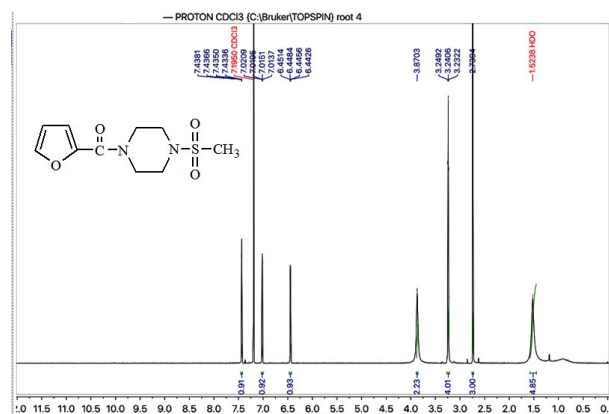
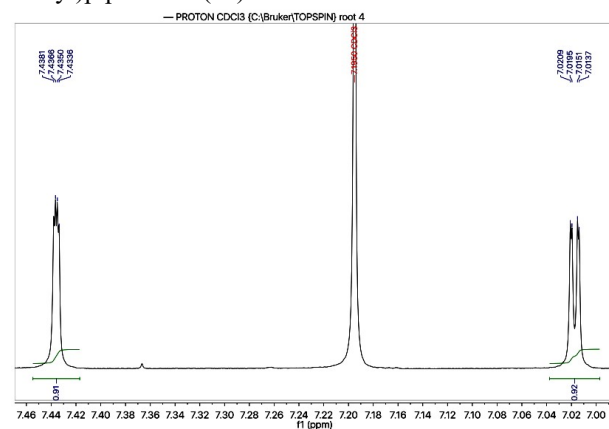
Compound	Inhibition (%)				
	<i>S. typhi</i> (-)	<i>E. coli</i> (-)	<i>P. aeruginosa</i> (-)	<i>B. subtilis</i> (+)	<i>S. aureus</i> (+)
3a	76.40±0.9	89.20±0.20	73.50±0.20	75.50±0.81	68.00±0.53
3b	68.75±0.95	72.35±1.14	60.57±0.43	70.55±0.95	67.15±0.95
3c	77.80±1.20	79.25±0.25	75.00±0.25	79.25±1.15	73.43±0.98
3d	75.85±0.35	74.80±0.20	68.21±0.86	78.70±1.20	64.86±0.05
3e	75.85±1.45	71.50±1.20	77.79±1.07	77.400±1.00	64.56±1.04
3f	74.05±1.55	74.30±0.50	65.36±0.35	75.85±0.53	64.64±1.00
3g	63.50±0.8	58.60±1.27	50.21±0.30	65.80±0.60	52.64±1.00
3h	78.45±0.65	77.00±1.27	65.93±0.60	76.55±1.25	67.29±0.44
3i	84.75±0.85	75.75±0.14	76.21±1.00	74.30±0.20	66.07±0.95
Ciprofloxacin	92.87±0.91	92.27±0.64	92.34±0.35	91.63±0.05	90.57±0.35

Table 2: Antibacterial activity (MIC, μM) of the synthesized compounds.

Compound	MIC (μM)				
	<i>S. typhi</i> (-)	<i>E. coli</i> (-)	<i>P. aeruginosa</i> (-)	<i>B. subtilis</i> (+)	<i>S. aureus</i> (+)
3a	9.26±0.05	7.89±0.79	9.02±0.20	8.01±0.33	10.37±0.51
3b	9.11±0.91	9.14±0.28	13.87±0.58	10.38±0.19	10.35±0.69
3c	9.24±0.90	8.27±0.73	9.26±0.37	8.05±0.10	10.38±0.38
3d	9.98±0.95	9.02±0.36	9.87±0.86	8.10±0.47	12.64±0.75
3e	9.93±0.05	10.48±0.58	8.11±0.47	8.39±0.21	12.58±0.83
3f	9.36±0.55	9.47±0.79	10.24±0.16	9.12±0.22	12.67±0.79
3g	11.57±0.90	14.36±0.47	17.63±0.85	12.06±0.58	17.46±0.38
3h	8.98±0.61	8.19±0.37	10.47±0.70	8.09±0.42	10.38±0.57
3i	8.10±0.85	9.46±0.43	9.74±0.36	10.21±0.57	10.97±0.63
Ciprofloxacin	7.28±0.11	7.31±0.14	7.22±0.76	7.29±0.05	7.86±0.42

Table 3: Hemolytic activity of the synthesized compounds.

Compound	Hemolytic activity
	% Hemolysis
3a	83.83
3b	70.88
3c	27.87
3d	56.27
3e	81.20
3f	86.32
3g	29.13
3h	82.85
3i	88.57
PBS	0.43
Triton	100

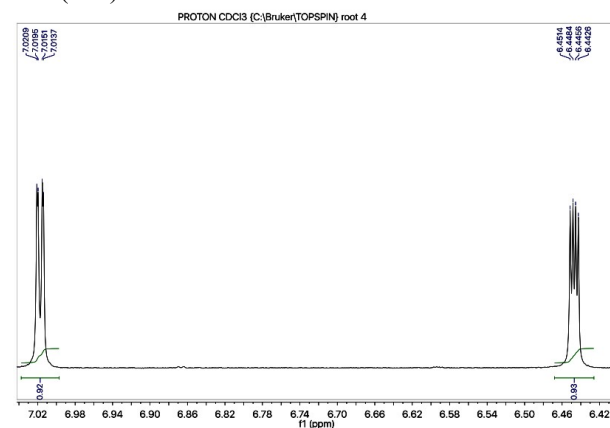
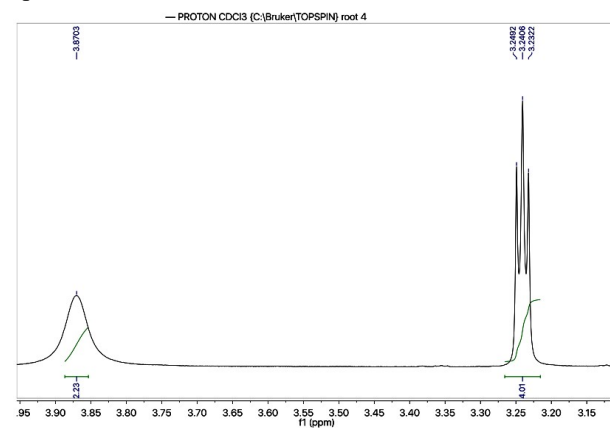
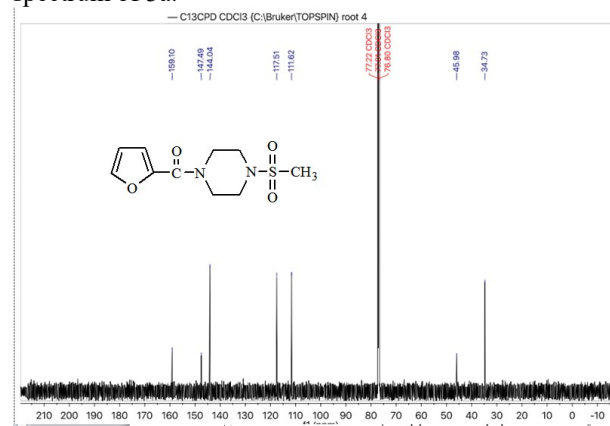
**Fig. 1a:** $^1\text{H-NMR}$ spectrum of 4-(methylsulfonyl)-1-(2-furoyl)piperazine (3a).**Fig. 1b:** Expanded downfield region of $^1\text{H-NMR}$ spectrum of 3a.

DISCUSSION

Chemistry

4-(Methylsulfonyl)-1-(2-furoyl)piperazine (3a) was purified as white amorphous solid and its molecular formula $\text{C}_{10}\text{H}_{14}\text{N}_2\text{O}_4\text{S}$ was ascertained by counting the number of protons in its $^1\text{H-NMR}$ spectrum (fig. 1a-d) and number of carbon resonances in its $^{13}\text{C-NMR}$

spectrum (fig. 2). The presence of prominent functionalities was also confirmed by IR absorption bands at 3027 (Ar C-H), 2836 (R C-H), 1603 (C=O), 1584, 1453 (C=C), 1323 (S=O), 1156 (C-O-C), 1078 (C-N-C), and 655 (C-S).

**Fig. 1c:** Expanded part of aromatic region of $^1\text{H-NMR}$ spectrum of 3a.**Fig. 1d:** Expanded part of aliphatic region of $^1\text{H-NMR}$ spectrum of 3a.**Fig. 2:** $^{13}\text{C-NMR}$ spectrum of 3a.

The singlet at δ 2.73 (s, 3H, CH_3 -1") was allotted to methyl group directly attached to sulfonyl group. The furan moiety showed three singlets, with one-proton

integration each, at δ 7.43 (dist.d, $J=0.9$ Hz, 1H, H-5), 7.02 (d, $J=2.6$ Hz, 1H, H-3) and 6.45 (dd, $J=1.8, 3.4$ Hz, 1H, H-4). The piperazine moiety presented one broad singlet and one triplet, for the overall eight-portons, resonating at δ 3.87 (br.s, 4H, CH₂-3' & CH₂-5') and 3.24 (br.t, $J=5.1$ Hz, 4H, CH₂-2' & CH₂-6'). The ¹³C-NMR spectrum (fig. 2) further augmented the structural interpretation. The carbon core of furoyl moiety was fully confirmed by five signals of 159.1 (C-6), 147.4 (C-2), 144.0 (C-5), 117.5 (C-4) and 111.6 (C-3). The four carbons of piperidine moiety were merged into one signal at δ 45.9 (C-2', C-3', C-5', C-6'), owing to usual ring flipping. Similarly, the appearance of a singlet at δ 34.7 (C-1") was rational for a methyl group attached to sulfonyl functionality. So, on the basis of cumulative spectral evidences the structure of 3a was affirmed as 4-(methylsulfonyl)-1-(2-furoyl)piperazine. Same protocol was followed for the structural characterization of other synthesized compounds.

Antibacterial activity

The synthesized compounds remained efficient inhibitors of all the considered bacterial strains. The lower MIC values, which are comparable to the reference, ciprofloxacin, supported better activity of these compounds. Against *S. typhi*, 4-(4-methoxy phenylsulfonyl)-1-(2-furoyl)piperazine (3i) remained the most active compound with MIC value of 8.10±0.85µM in comparison of that of reference, 7.28±0.11µM. In broader interpretation, only 3g was the less active against this strain. 4-(Methylsulfonyl)-1-(2-furoyl)piperazine (3a) was the most active compound against *E. coli*, with MIC value of 7.89±0.79µM, relative to MIC of reference, 7.31±0.14µM. Against *P. aeruginosa*, 4-(4-methylphenyl sulfonyl)-1-(2-furoyl)piperazine (3e) was the most active one with MIC value of 8.11±0.47µM. The reference was having MIC 7.22±0.76µM in this case. Against *B. subtilis*, 4-(methylsulfonyl)-1-(2-furoyl)piperazine (3a) was the most superb one with MIC value of 8.01±0.33 µM in comparison of the reference having value of 7.29±0.50 µM. Against *S. aureus*, 4-(2-naphthylsulfonyl)-1-(2-furoyl)piperazine (3b) was identified as best active molecule bearing MIC value of 10.35±0.69µM, relative to the reference exhibiting MIC value of 7.86±0.82µM. In general, all the synthesized molecules exhibited convincing antibacterial inhibition potential, except 3g.

Hemolytic activity

Two compounds, 3c bearing phenyl and 3g bearing 4-acetylphenyl groups, exhibited very mild hemolytic activity with % lysis of 27.87 and 29.13%, respectively, relative to the Triton-X-100 (% lysis 100). Highest % lysis was shown by 3i (% lysis 88.57) yet it is much lower than positive control Triton-X-100. So, in general it is pertinent to say that the studied molecules are not toxic and might be utilized as safe therapeutic agents.

CONCLUSION

It was concluded from the investigated study that targeted sulfonamides were successfully synthesized in good yields by a benign and cost effective method. Most of the synthesized compounds exhibited remarkable antibacterial potentials with mild cytotoxicity. So, such molecules might serve as valued lead compounds in drug discovery and development program.

REFERENCES

- Abbasi MA, Zeb A, Aziz-ur-Rehman, Siddiqui SZ, Shah SAA, Shahid M and Fatima H (2020^a). Synthesis, bacterial biofilm inhibition and cytotoxicity of new *N*-alkyl/aralkyl-*N*-(2,3-dihydro-1,4-benzodioxin-6-yl)-4-nitrobenzenesulfonamides. *Pak. J. Pharm. Sci.*, **33**(1): 41-47.
- Abbasi MA, Irshad M, Aziz-ur-Rehman, Siddiqui SZ, Nazir M, Shah SAA, Shahid M (2020^b). Synthesis of promising antibacterial and antifungal agents: 2-[[4-(4-Chlorophenyl)sulfonyl](2,3-dihydro-1,4-benzodioxin-6-yl)amino]-*N*-(un/substituted-phenyl)acetamides. *Pak. J. Pharm. Sci.* **33**(5): 2161-2170.
- Apaydin S, and Török M (2019). Sulfonamide derivatives as multi-target agents for complex diseases. *Bioorg. Med. Chem. Lett.*, **29**(16): 2042-2050.
- Bruder P, Grimstvedt A, Mejdell T and Svendsen HF (2011). CO₂ capture into aqueous solutions of piperazine activated 2-amino-2-methyl-1-propanol. *Chem. Eng. Sci.*, **66**(23): 6193-6198.
- Clercq, ED (2001). New developments in anti-HIV chemotherapy. *Curr. Med. Chem.*, **8**(13): 1543-1572.
- Dou D, He G, Mandadapu SR, Aravapalli S, Kim Y, Chang K and Groutas WC (2012). Inhibition of noroviruses by piperazine derivatives. *Bioorg. Med. Chem. Lett.*, **22**(1): 377-379.
- Eloff JN (1998). A sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria. *Planta Med.*, **64**(8): 711-713.
- El-Sayed NS, El-Bendary RE, El-Ashry SM and El-Kerdawy MM (2011). Synthesis and antitumor activity of new sulfonamide derivatives of thiazolo[3,2-a]pyrimidines. *Eur. J. Pharm. Sci.*, **46**(9): 3714-3720.
- Hassan AU, Sumrra SH, Raza MA, Zubair M, Zafar MN, Mughal EU, Nazar MF, Irfan A, Imran M and Assiri MA (2021). Design, facile synthesis, spectroscopic characterization and medicinal probing of metal-based new sulfonamide drugs: A theoretical and spectral study. *Appl. Organomet. Chem.*, **35**(1): 6054-6071.
- Jalageri MD, Puttaiahgowda YM and Hariprasad (2019). Design and antimicrobial activity of piperazine polymer nanocomposite. *Materials Today: Proceedings.*, **15**(2): 262-267.
- Kaspady M, Narayanaswamy VK, Raju M and Rao GK (2009). Synthesis, antibacterial activity of 2,4-

- disubstituted oxazoles and thiazoles as bioisosteres. *Lett. Drug Des. Discov.*, **6**(1): 21-28.
- Mirian M, Zarghi A, Sadeghi S, Tabaraki P, Tavallae M, Dadrass O and Sadeghi-Aliabadi H (2011). Synthesis and cytotoxic evaluation of some novel sulfonamidederivatives against a few human cancer cells. *Iran. J. Pharm. Res.*, **10**(4): 741-748
- Mohs, RC and Greig N (2017) Perspective drug discovery and development: Role of basic biological research. *Alzheimer's & Dementia Translational Research & Clinical Interventions*. **3**(4): 651-657.
- Mun J, Jabbar AA, Devi NS, Yin S, Wang Y, Tan C, Culver D, Culver JP, Meir EGV and Goodman, M. M. (2012). Design and in vitro activities of *N*-alkyl-*N*-[(8-*R*-2,2-dimethyl-2H-chromen-6-yl)methyl] heteroarylsulfonamides, novel, small-molecule hypoxia inducible factor-1 (HIF-1) pathway inhibitors and anticancer agents. *J. Med. Chem.*, **55**(15): 6738-6750.
- Ozbek N, Ozdemir OO, Altun AF and Sahin E (2019). Sulfonamide-derived hydrazone compounds and their Pd (II) complexes: Synthesis, spectroscopic characterization, X-ray structure determination, in vitro antibacterial activity and computational studies. *J. Mol. Str.*, **15**(2): 707-719.
- Powell WA, Catranis CM and Maynard CA (2000). Design of self-processing antimicrobial peptides for plant protection. *Lett. Appl. Microbio.*, **31**(2): 163-168.
- Rehman H, Qadir A, Ali Z, Nazir S, Zahra A and Shahzady TG (2017). Synthesis and characterization of novel sulfonamides derivatives and their antimicrobial, antioxidant and cytotoxicity evaluation. *Bull. Chem. Soc. Ethiop.*, **31**(3): 491-498.
- Rodríguez A, Villegas E, Montoya-Rosales A, Rivas-Santiago B, Corzo G (2014). Characterization of antibacterial and hemolytic activity of synthetic Pandinin 2 variants and their inhibition against *Mycobacterium tuberculosis*. *PLoS One.*, **9**(7): 101742-101753
- Root H, Daniels L, Marx A, Bartelt LA, Lachiewicz AM, Duin DV (2021). Sulfonamides without trimethoprim in the treatment of *Nocardia* infections. *Transpl. Infect. Dis.*, **23**(1):13452-13457.
- Saeedi M, Goli F, Mahdavi M, Dehghan G, Faramarzi MA, Foroumadi A and Shafiee A (2014). Synthesis and biological investigation of some novel sulfonamide and amide derivatives containing coumarin moieties. *Iran. J. Pharm. Res.*, **13**(3): 881-892.
- Shahid M, Bukhari SA, Gul Y, Munir H, Anjum F, Zuber M, Jamil T and Zia KM (2013). Graft polymerization of guar gum with acryl amide irradiated by microwaves for colonic drug delivery. *Inter. J. Bio. Macromol.*, **62**(2): 172-179.
- Sharma P and Sharma JD (2001). *In vitro* hemolysis of human erythrocytes by plant extracts with antiplasmodial activity. *J. Ethnopharmacol.*, **74**(3): 239-243.
- Tang YL, Li YK, Li MX, Gao H, Yang XB and Mao ZW (2020). Synthesis of new piperazine substituted chalcone sulphonamides as antibacterial agents. *Curr. Org. Synth.* **17**(2): 136-143.
- Thach TD, Nguyen TMT, Nguyen TAT, Dang CH, Luong TB, Dang VS, Banh KS, Luc VS, Nguyen TD (2021). Synthesis and antimicrobial, antiproliferative and anti-inflammatory activities of novel 1,3,5-substituted pyrazoline sulphonamides, *Arab. J. Chem.*, **14**(11): 103408-103419
- Yamaguchi I, Kado A, Fukuda T, Fukumoto H, Yamamoto T and Sato M (2010). Ionic polymers and oligomers with expanded π -conjugation system derived from through-space interaction in piperazinium ring. *Eur. Polym. J.*, **46**(5): 1119-1130.
- Yang CR, Zang Y, Jacob MR, Khan SI, Zhang YJ and Li XC (2006). Antifungal activity of C-27 steroidal saponins. *Antimicrob. Agents Chemother.*, **50**(5): 1710-1714.
- Zoumpoulakis P, Camoutsis C, Pairas G, Sokovic M, Glamoclija J, Potamitis C and Pitsas A (2012). Synthesis of novel sulfonamide-1,2,4-triazoles, 1,3,4-thiadiazoles and 1,3,4-oxadiazoles, as potential antibacterial and antifungal agents. Biological evaluation and conformational analysis studies. *J. Bioorg. Med. Chem.*, **20**(4): 1569-1583.