# Pharmacological effects of *Raphanus caudatus* in inflammation and hematopoitic system: an *in vitro* and *in vivo* investigation

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Abstract: The pods of Radish are known as *Raphanus caudatus that* belongs to the family *Brassicaceae*. They are commonly known as Mungra or Sungra in Pakistan, while the common English name for this species is Rat-tailed radish. This variety of radish is unique and less tested for pharmacological as well as toxicological potential. In the current research, the ethanol extract of pods was assessed for anti-inflammatory potential *in vitro* and *in vivo*. Furthermore the effect of plant on hematological parameters was also investigated. For *in vitro* study, luminol-enhanced chemiluminescence method was used while *in vivo* study was carried out via Acetic acid- induced Paw Edema Test in wistar rats. The extract of *Raphanus caudatus* indicated significant anti-inflammatory effects regarding *in vitro* assay. Administered tested doses (250mg, 500mg and 1000mg/kg) of plant extract showed significant reduction in rat's paw but highest *in vivo* anti-inflammatory effect was observed at the dose of 1000mg/kg. Moreover, in the case of hematological study, noticeable elevation of white blood cell count was observed at 500 and 1000 mg/kg. However the number of platelets was reduced in dose dependent manner.

**Keywords**: Raphanus caudatus, in vitro, in vivo, anti-inflammatory, hematology.

#### INTRODUCTION

The significance of vegetables cannot be ignored as they have very important role in our life not only as food materials but they are used in the treatment and inhibition of different diseases. These vegetables provide major source of amino acids, vitamins, antioxidants, minerals, and fibers so they are beneficial in the development and restoration of body deficiencies (Boeing, Bechthold *et al.* 2012, Slavin and Lloyd 2012).

Raphanus sativus L. Var. caudatus are edible and green in color (fig. 1), processes beneficial properties for health. It grows annually and sometimes biennially, seed pods which are formed from seeds lengthen about 30-90 cm tall and can grow up to 5 feet in height and can spread up to 2 feet (Pocasap, Weerapreeyakul et al. 2013). Commonly in Pakistan and India it is known as Mungraa or Sungraa. The seed pod part of radish is available in Pakistan from the months of November till March and is consumed in delicious dishes (Khare 2007). Radish is a rich source of diversified types of flavonoids such as apigenin, quercetin, kaempferol, luteolin and myricetin (Lugast and Hovari 2000). Other studies also reports the presence of phenolic content in Raphanus sativus (Jamuna, Ramesh et al. 2015, Kim, Moon et al. 2015). Moreover different phenolic compounds identified in radish are reported as anti-inflammatory such as Luteolin Kaempferol, Kaempferol-7-O-rhamnoside, Kaempferol-3glucoside-7-rhamnoside and Kaempferol-7-glucoside-3rhamnoside (Gutiérrez and Perez 2004, Sham, Yuen et al. 2013). The pods of Radish are good source of Vitamin C,

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Vitamin B9, pyridoxine, Vitamin B2 and Vitamin E (Songsak and Lockwood 2002, Pocasap, Weerapreeyakul *et al.* 2013).

Previously various pharmacological activities of radish pods were explored by our research project (Younus and Siddiq 2017, Siddiq and Younus 2018, Younus and Siddiq 2022). In the current study, anti-inflammatory potential of this vegetable along with its effect on hematological parameters were identified by employing *in vitro* and *in vivo* experimental designs.

#### MATERIAL AND METHODS

#### Plant material

Fresh pods (10kg) of *Raphanus sativus* L.Var. *caudatus* were procured from Karachi, identified by Associate Professor of University of Karachi Department of Pharmacognosy, Dr. Mohtashim and specimen with voucher number: RSP-01-14/17 was deposited to Department of Pharmacognosy. Pods of the plant were washed using double distilled water thoroughly after which dried in shade at room temperature and saved in properly labeled air tightcontainers.

#### Plant material extraction

Extract of the plant material was obtained at the temperature of 60°C using Ethanol (Merck) (1:50, w/v) from soxhlet apparatus (HMFT-5/63, England)(Okoduwa, Umar *et al.* 2016). The extraction process was performed repetitively until exhaustion (48h) and then extract obtained was filtered with autoclaved Whatman No. 1 filter paper. After filtration process of drying of filtrate was performed in a rotary evaporator (Buchi R-200,

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Switzerland) under controlled pressure and temperature of 45°C to obtain semisolid mass.

# Immunomodulatory/anti-inflammatory activity – in vitro study

Chemiluminescence assay

Evaluation of immunomodulatory activity of RCE was performed with Luminol-amplified chemiluminescence method (Helfand, Werkmeister et al. 1982). The Test was carried out in white 96-well plates (Costar, NY, USA). A25 µl cell suspension of whole blood (diluted) in Hanks Balanced Salt Solution, containing CaCl<sub>2</sub> and MgCl<sub>2</sub> (HBSS++) (Sigma, St. Louis, USA) was incubated with 25 ul of RCE in HBSS++ at three different concentrations i.e. 1, 10 and 100µg/ml. Medium with HBSS++ and cells only were kept as control wells. Incubation was performed at 37°C in the thermostat chamber of luminometer [Labsystems, Helsinki, Finland] for 15 minutes. Afterward, SOZ (serum opsonized zymosan) [Fluka, Buchs, Switzerland] and luminol (Research Organics, Cleveland, OH, USA), were (25µl) separately transferred to each well; however in blank wells contained with HBSS++. The ROS levels in term of RLU (relative light units) were determined by using luminometer and results were expressed as IC50 (50% inhibitory concentration). The tests were conducted in triplets.

#### In vivo study

Animals

All the animals were procured from Dow University of Health Sciences, Karachi. For anti-inflammatory activity, wistar rats weighing 200-250g and for biochemical testing on hematological parameters, albino rabbits weighing 1200-1800 g were used in the present study. All the animals were of both genders. Transparent cages were used for housing of rabbits which were housed individually in each cage while mice were housed in group of 3-6 per cage, under controlled conditions, temperature and humidity was (23±2°C), (50 to 60%) respectively in a 12/12h light and dark cycle. All Animals had access of water and fed with ad labitum. Prior to the initiation of the study all animals were observed for 1 week for acclimatization purpose to the laboratory environment and those were excluded having any infections.

# Anti- inflammatory activity of Raphanus caudtus extract Protocol of dosing

For anti-inflammatory activity of radish pods in vivo evaluation potential was done by acetic acid induced paw edema method in albino wistar rats. There were 5 groups of animals (n=10 each). All drugs were dissolved in distilled water and administered via oral route. The standard drug in the form of tablets was purchased from local market.

#### Acetic acid- induced paw edema test

Firstly animals of all groups were parenterally administered with 1% w/v acetic acid (0.1ml) in the right hind paw of rat's sub-plantar tissue (Bachhav, Gulecha *et al.* 2009) to induce edema. After 30 min administration of acetic acid all animals were treated orally with normal saline, standard drug (acetylsalicylic acid) and RCE (250, 500 and 1000mg/kg) respectively.

In the current study, Plethysmometer (UGO Basile, Italy) is used for the measurement of paw volume (in term of ml). The readings were recorded at "0" time (baseline), following acetic acid treatment and observed hourly till 4 hours after administration of edematogenic agent.

Edematous paw reduction was calculated by using following formula in term of percentage(Gupta, Mazumder *et al.* 2005, Rao and Najam 2016), Khobragade *et al.*, 2012).

(%) Inhibition of edematous paw = 
$$\frac{C - T}{C} \times 100$$

Where.

C = Average increase in paw volume of control rat at given time T= Average increase in paw volume of treated rat at given time

#### Hematological study

Hematological parameters were observed in rabbits divided in to three groups (each contains 10 animals) i.e. RCE I, II and III (250, 500 and 1000mg/kg) were administered once daily for continuous 60 days. After dosing protocol of 60 days, 2 ml blood was collected in EDTA.K3 tubes for hematological screening. Blood samples that were collected in siliconized glass tubes (Heyns, van den Berg *et al.* 1981) investigated for hematological parameters (RBC, WBC, platelet count and hemoglobin level) by automatic Humacount plus Germany.

#### Ethical approval

All the research protocol, procedures, euthanasia as well as ethical clearance were approved by the BASR (Board of Advanced Studies & Research), University of Karachi (BASR number 02419/Pharm). All the selected animals were handled using ethical principles of research. Specifications described in Helsinki Resolution 1964 were adopted for handling of animals in the current research.

#### STATISTICAL ANALYSIS

All data analysis and statistical procedures were performed using Statistical Package for the Social Sciences (SPSS) version 20. The results were presented as mean + standard error of mean. The groups were compared by One - way ANOVA (Analysis of variance) followed by post hoc Tukey's Honest Significant Difference (HSD) test to find out statistical significance.

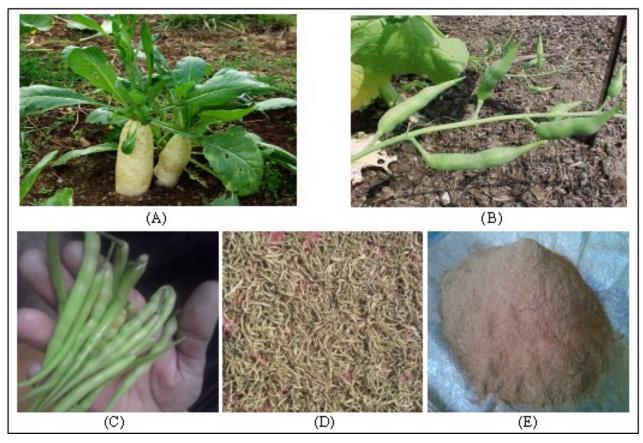


Fig. 1: Raphanus sativus L. (A) Roots (B), (C) Fresh Pods (D) Dried pods (E) Powder of dried Pods (imageswere taken from current study and flora of Pakistan).

Table 1: Dosing Protocol for In vivo Study

Groups	Treatment	Dose (mg/kg) p.o	
Control	Normal saline	1 ml/kg	
Standard	Aspirin*	300	
Test group I	RCE-A	250	
Test group II	RCE-B	500	
Test group III	RCE-C	1000	

<sup>\*</sup>used as standard drug for anti-inflammatory activity.

Results obtained from current data were regarded as significant and highly significant at p<0.05 and p<0.005 respectively.

#### **RESULTS**

### In vitro Anti-inflammatory / Immunomodulatory activity The immunomodulatory activity of ethanol extract of

The immunomodulatory activity of ethanol extract of *Raphanus caudatus* showed significant results with more  $IC_{50}$  value (37.8  $\pm$  2.1) as compare to standard IC50 (11.3  $\pm$  1.89) as shown in table 2.

#### In vivo anti- inflammatory activity

The anti-inflammatory effect of RCE was determined by acetic acid induced paw edema in rats at three different doses and acetylsalicylic acid is shown in table 3. The

revealed anti-inflammatory activity of RCE showed time and dose dependent inhibition in term of (%) inhibition of paw edema and displacement in ml  $\pm$  SEM.

The results showed that given extract of RCE Aat 250mg/kg revealed significant effect (p<0.05) on paw edema i.e. (%) inhibition of paw edema after 1h and 2h of acetic acid administration was 19.04% and 18.36% as compare to control group. The significant reduction (p<0.005) of rat paw edema against acetic acid administration was observed in time dependent manner at 3<sup>rd</sup> and 4<sup>th</sup>hr following administration.

Animal treated with RCE B extract presented prominent reduction in edematous paw volume induced by acetic acid in rat observed volume 1h post treatment was  $3.5\pm0.17$ ml (16.66%) in comparison of saline administered group where displaced volume was i.e.  $4.2\pm0.10$ ml.

**Table 2**: Immunomodulatory activity of *Raphanus* caudatus in vitro by Chemiluminiscence assay

Immunomodulatory study			
$IC_{50} \pm SEM (\mu g/ml)$			
RCE	$37.8 \pm 2.1$		
Ibuprofen	$11.3 \pm 1.89$		

The highest tested dose in this study i.e. RCE-C administered in animals revealed prominent anti-inflammatory activity in comparison to control group animals which is approximate of standard drug acetylsalicylic acid The results indicate that animals having drug acetylsalicylic acid exhibits potent anti-inflammatory effect by decrement in rat paw edema after the acetic acid administration from 1 to 4h.

## Effect of Raphanus caudatus on hematological parameters

The effect of RCE on hematological parameter (RBCs, WBCs and Platelets count) at three different doses is showed in table 4. After 60 days dosing protocol there was no significant RBCs count alterations in all three group treated with RCE (A, B and C). With respect to control in the group of RCE-A effect on WBCs count results were statistically insignificant. While group

administered with RCE-B and RCE-C, WBCs count were highly significant increase (p<0.05, p<0.005) by (5.2±0.21\*), (7.2±0.18\*\*) respectively. The results of *Raphanus caudatus* on animals having RCE-A, platelet count was significant (p<0.05) in comparison with control group. Further, the platelet count in animals treated with RCE-B and RCE-C revealed highly significant reduction (p<0.005) when compared with control group. But effect on Hemoglobin levels of RCE (A, B and C) showed no significant effect in contrast to control group

#### **DISCUSSION**

Raphanus caudatus is known for its health benefits. In this study the pharmacological effects of this variety of radish have been identified in inflammation / immunomodulation. Any modification in immune system by any stimuli is called Immunomodulation. It is required in condition of immunodeficiency to combat with number of immune disorders (Gea-Banacloche 2006). In earlier study it has been showed that radish have immunomodulatory potential, which has been observed by the aqueous extract of Raphanus niger as it showed increase in survival rate and also provide protection against experimental models of influenza infection in mice (Prahoveanu and Esanu 1986).

It is approved that ROS are produced after the activation PMNs and formation of ROS is known as oxidative burst.

**Table 3**: Effect of *Raphanus caudatus* and Aspirin on percent inhibition of paw edema in Acetic acid-induced Paw Edema Test

Treatment	Displacement in 'ml'		Displacement in 'ml' and (percent inhibition) after acetic acid treatment			
Groups	Pre-drug	Immediately after acetic acid administration 0 h	1 h	2 h	3 h	4 h
Control	2.7±0.12	2.9±0.13	4.2±0.10	4.41±0.18	4.9±0.18	4.84±0.21
Aspirin	2.6±0.14	.14 2.9±0.16	3.15±0.14**	3.19±0.14**	3.28±0.10**	3.34±0.12**
(300 mg/kg)	2.0±0.14		(25.0)	(27.66)	(33.06)	(31)
RCE-A	3.1±0.15	2.4±0.15	3.4±0.18*	3.6±0.16*	3.5±0.15**	3.62±0.11**
(250 mg/kg)	3.1±0.13	2.4±0.13	(19.04)	(18.36)	(28.57)	(25.2)
RCE -B	2.9±0.15	3.1±0.12	3.5±0.17*	3.25±0.13**	3.43±0.18**	3.52±0.20**
(500 mg/kg)	2.9±0.13	3.1±0.12	(16.66)	(26.30)	(30)	(27.27)
RCE-C	3.2±0.11	3.0±0.14	3.36±0.11**	2.73±0.18**	2.69±0.21**	3.15±0.25**
(1000 mg/kg)	3.2±0.11	3.0±0.14	(20.0)	(38.09)	(45.10)	(35)

The values in brakettes indicate percentage edema inhibition. n=10 per group, Values are Mean  $\pm$  Std. Error of Mean, RCE: Raphanus caudatus Extract, \*p  $\leq$  0.05 significant versus control, \*\*p  $\leq$  0.005 highly significant versus control

 Table 4: Effect of Raphanus caudatus on Hematological Parameters

Treatment Groups	RBCs (x10 <sup>6</sup> /µl)	WBCs $(x10^3/\mu l)$	Platelets (x10 <sup>3</sup> /μl)	Hb (gm/dl)
Control	4.1±0.19	4.4±0.22	425±31.16	$9.3 \pm 0.34$
RCE-A (250 mg/kg)	4.2±0.2	4.2±0.21	375±15.16*	8.9±0.15
RCE-B (500 mg/kg)	3.9±0.17	5.2±0.21*	352±39.91**	9.9±0.55
RCE-C (1000 mg/kg)	4.1±0.14	7.2±0.18**	212±22.81**	9.1±0.26

n =10 per group, Values are Mean  $\pm$  Std. Error of Mean, RCE: Raphanus caudatus Extract, \*p  $\leq$  0.05 significant versus control, \*\*p < 0.005 highly significant versus control

It is fact that these ROS provide protection against pathogenic microorganisms, but it also causes tissue damage which ultimately lead to progression of inflammatory disorders which might be reason of various diseases (Johansen, Harris *et al.* 2005). Chemiluminescence assay is based on the luminescent capacity of luminol by the attacking of free radicals in the sample (Roginsky and Lissi 2005).

This study revealed that RCE significantly have immunomodulatory/anti-inflammatory effect against ROS in vitro testing by chemiluminescence assay. Activation of antioxidant enzymes and inhibitory potential against ROS has been studied in another study by Tunisian radish extract (Salah-Abbès *et al.* 2008). Lately Salah-Abbès and co-workers also stated about the protective effects of root extract of *Raphanus sativus* in 2010 while immunotoxicity was induced by high dose of zearalenone. Fruits and Vegetables are the rich source of alkaloids, saponin and phenols (Liu 2003 and Oz 2017) that have very important effect in inhibition of oxidation process that could be attributed to observed effects in RCE.

As we all know that inflammation is protective mechanism of body and in our current study when inflammation was induced by acetic acid induced rat paw edema model the results of table 3 findings showed that RCE have significant anti-inflammatory effect (Libby 2007). The inflammatory mediators majorly effect in inflammation are released in two stages.

Initially in two hours of inflammatory mediators like histamine and serotonin are released while in second stage prostaglandins, proteases and kinins are released after 3-4 hours of acetic acid injection which plays major role in inflammation (Lomba et al., 2017). After the dosing protocol all tested doses (250, 500 and 1000mg/kg) of RCE, we have found significant reduction in rat's paw while the highest percentage of paw edema inhibition 45.01% was observed at the dose of 1000 mg/kg which is greater than our standard drug acetylsalicylic acid at same time i.e. 33.06%. So it indicates that RCE may act particularly via inhibition of prostaglandin synthesis. Kim et al. (2015 and 2014) found in their studies that seeds of Raphanus caudatus have some anti- inflammatory derivatives i.e. phenylpropanoid sucrosides methylthio-butanyl which were observed by the inhibition of nitric oxide during in vitro anti- inflammatory activity (Kim et al. 2014 and 2015). In the other study of water extract of Raphanus caudatus seeds alleviate intestinal inflammation in trinitrobenzenesulphonic acid and dextran sodium sulfate -induced colitis in rats. The occurrence of sinapic acid was attributed to this antiinflammatory effect (Kook et al. 2014 and Cho et al., 2016).

The occurrence of flavonoids has been reported in the pods of radish (Takaya et al., 2003, Beevi et al., 2012) similarly in our study the anti-inflammatory activity of RCE, this might be a reason of this effect and support the folk use of Raphanus sativus in these disorder (B Aggarwal, Prasad et al. 2011). Furthermore, Radish is a great source of vitamin C which is well reported in different studies of anti-inflammatory and antioxidant activity (Peters, Anderson et al. 2001, Padayatty, Katz et al. 2003).

This study provides evidence that that leukocytes level was increased significantly by RCE at the completion of 500 and 1000mg/kg. Leukocytes have major role in defensive system of body. Increased leukocyte count supports the immunosuppression and healing of wounds. Though elevated leukocytes are the markers of blood related diseases and long term cancer like leukemia. Since it may assumes that the rich amount of alkaloids, flavonoids, terpenoid and isothiocyante compounds in *Raphanus caudatus* indicates enhance leukocyte levels in the current study and it has been supported by the Thejass and Kuttan (2007).

This study of RCE showed a remarkable reduction in number of platelet in dose dependent manner after longterm administration. In the current study, the platelet inhibiting activity of Radish pods is an interesting outcome. As discussed previously that Radish pods have high amount of alkaloids and isothiocyanate compound, especially sulforaphene and sulforaphane which possess anti-platelet activity that has been reported in several studies (Chuang et al., 2013, Fuentes and Palomo 2014, Jayakumar et al., 2016). However sulforaphane antiplatelet activity is reported in different studies which is due to stimulation of adenylate cyclase, glycoprotein IIb/IIIa as well as thromboxane A2 (Jayakumar et al., 2013, Oh et al., 2013, Ku and Bae 2014). The antiplatelet activity occurs due to the presence of alkaloids which is already reported in different studies (Chen et al., 1996). Therefore, the occurrence of natural compounds in Radish pods might be attributed to its platelet inhibiting activity in this study. The antiplatelet activity of Radish pods could have a protective effect in coronary artery diseases and thrombotic illnesses and caution must be taken in the usage before surgery.

#### **CONCLUSION**

Based on the results obtained from this study, it is concluded that *Raphanus caudatus* processes significant pharmacological potential in inflammation and hematological parameters. The plant produced significant anti-inflammatory activity *in vitro* and *in vivo*. This might be attributed because of the presence of flavonoids and vitamin C. Moreover, a statistically significant raised level of leukocytes at 500 and 1000 mg/kg with a dose-

dependent reduction in platelets count was prominent. It is recommended to carry out further study to explore the mechanism of observed anti-inflammatory and hematological potential.

#### REFERENCES

- Aggarwal B, Prasad S, Reuter S, Kannappan R, Yadav VR, Park B, Kim JH, Gupta SC, Phromnoi K and Sundaram C (2011). Identification of novel anti-inflammatory agents from Ayurvedic medicine for prevention of chronic diseases: "Reverse pharmacology" and "bedside to bench" approach. *Curr. Drug Targets*, **12**(11): 1595-1653.
- Bachhav R, Gulecha V and Upasani C (2009). Analgesic and anti-inflammatory activity of *Argyreia speciosa* root. *Indian J. Pharmacol.*, **41**(4): 158.
- Beevi SS, Mangamoori LN and Gowda BB (2012). Polyphenolics profile and antioxidant properties of *Raphanus sativus* L. *Nat. Prod. Res*, **26**(6): 557-563.
- Boeing H, Bechthold A, Bub A, Ellinger S, Haller D, Kroke A, Leschik-Bonnet E, Müller MJ, Oberritter H and Schulze M (2012). Critical review: Vegetables and fruit in the prevention of chronic diseases. *Eur. J. Nutr.*, **51**(6): 637-663.
- Chen KS, Ko FN, Teng CM and Wu YC (1996). Antiplatelet and vasorelaxing actions of some benzylisoquinoline and phenanthrene alkaloids. *Nat. Prod. Res.*, **59**(5): 531-534.
- Choi KC, Cho SW, Kook SH, Chun SR, Bhattarai G, Poudel SB, Kim MK, Lee KY and Lee JC (2016). Intestinal anti-inflammatory activity of the seeds of *Raphanus sativus* L. in experimental ulcerative colitis models. *J. Ethnopharmacol.*, **179**: 55-65.
- Chuang WY, Kung PH, Kuo CY and Wu CC (2013). Sulforaphane prevents human platelet aggregation through inhibiting the phosphatidylinositol 3-kinase/Akt pathway. *Thromb Haemost.*, **109**(6): 1120-1130.
- Fuentes E and Palomo I (2014). Antiplatelet effects of natural bioactive compounds by multiple targets: Food and drug interactions. *J. Funct. Foods*, **6**: 73-81.
- Gea-Banacloche JC (2006). Immunomodulation. Principles of Molecular Medicine, Springer, pp.893-904.
- Gupta M, Mazumder U, Kumar RS, Gomathi P, Rajeshwar Y, Kakoti B and Selven VT (2005). Anti-inflammatory, analgesic and antipyretic effects of methanol extract from *Bauhinia racemosa* stem bark in animal models. *J. Ethnopharmacol.*, **98**(3): 267-273.
- Gutierrez, RMP and RL Perez (2004). *Raphanus sativus* (Radish): Their chemistry and biology. *Sci. World J.*, 4: 811-837.
- Helfand SL, Werkmeister J and Roder JC (1982). Chemiluminescence response of human natural killer cells. I. The relationship between target cell binding,

- chemiluminescence and cytolysis. *J. Exp. Med.*, **156**(2): 492-505.
- Heyns A, van den Berg D, Kleynhans P and du Toit P (1981). Unsuitability of evacuated tubes for monitoring heparin therapy by activated partial thromboplastin time. *J. Clin. Pathol.*, **34**(1): 63-68.
- Jamuna KS, Ramesh CK, Mahmood R, Pallavi M and Rao SJA (2015). Effect of different extraction methods on total phenolic content and antioxidant activities of Raphanus sativus. *Int. J. Bioassays*, 4(12): 4653-4657.
- Jayakumar T, Chen WF, Lu WJ, Chou DS, Hsiao G, Hsu CY, Sheu JR and Hsieh CY (2013). A novel antithrombotic effect of sulforaphane via activation of platelet adenylate cyclase: *ex vivo* and *in vivo* studies. *J. Nutr. Biochem,* **24**(6): 1086-1095.
- Jayakumar T, Yang CH, Geraldine P, Yen TL and Sheu JR (2016). The pharmacodynamics of antiplatelet compounds in thrombosis treatment. *Expert Opin. Drug Metab. Toxicol.*, **12**(6): 615-632.
- Johansen JS, Harris AK, Rychly DJ and Ergul A (2005). Oxidative stress and the use of antioxidants in diabetes: linking basic science to clinical practice. *Cardiovasc. Diabetol.*, **4**(1): 5.
- Khare C (2007). Indian Medicinal Plants: An illustrated dictionary Library of congress Control Number: 2007922446 Springer-Verlag Berlin, Heidelberg, Springer Science+ Business Media, LLC.
- Kim KH, Kim CS, Park YJ, Moon E, Choi SU, Lee JH, Kim SY and Lee KR (2015). Anti-inflammatory and antitumor phenylpropanoid sucrosides from the seeds of Raphanus sativus. *Bioorganic Med. Chem. Lett.*, **25**(1): 96-99.
- Kim KH, Moon E, Kim SY, Choi SU, Lee JH and Lee KR (2014). 4-Methylthio-butanyl derivatives from the seeds of *Raphanus sativus* and their biological evaluation on anti-inflammatory and antitumor activities. *J. Ethnopharmacol.*, **151**(1): 503-508.
- Kim KH, Moon E, Lee SR, Park KJ, Kim SY, Choi SU and Lee KR (2015). Chemical constituents of the seeds of *Raphanus sativus* and their biological activity. *J. Braz. Chem. Soc.*, **26**(11): 2307-2312.
- Kook SH, Choi KC, Lee YH, Cho HK and Lee JC (2014). *Raphanus sativus* L. seeds prevent LPS-stimulated inflammatory response through negative regulation of the p38 MAPK-NF-κB pathway. *Int. Immuno-pharmacol.*, **23**(2): 726-734.
- Ku SK and Bae JS (2014). Antithrombotic activities of sulforaphane via inhibiting platelet aggregation and FIIa/FXa. *Arch. Pharm. Res.*, **37**(11): 1454-1463.
- Libby P (2007). Inflammatory mechanisms: The molecular basis of inflammation and disease. *Nutr. Rev.*, **65**(Suppl 3): S140-S146.
- Liu RH (2003). Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. *Am. J. Clin. Nutr.*, **78**(3): 517S-520S.
- Lomba LA, Vogt PH, Souza VE, Leite-Avalca MC, Verdan MH, Stefanello MEA and AR Zampronio

- (2017). A naphthoquinone from *Sinningia canescens* inhibits inflammation and fever in mice. *Inflammation*, pp.1-11.
- Lugast A and Hovari J (2000). Flavonoid aglycons in foods of plant origin I vegetables. *Acta Aliment.*, **29**(4): 345-352.
- Oh CH, Shin JI, Mo SJ, Yun SJ, Kim SH and Rhee YH (2013). Antiplatelet activity of L-sulforaphane by regulation of platelet activation factors, glycoprotein IIb/IIIa and thromboxane A2. *Blood Coagul. Fibrinolysis*, **24**(5): 498-504.
- Okoduwa SIR, Umar IA, James DB, Inuwa HM and Habila JD (2016). Evaluation of extraction protocols for anti-diabetic phytochemical substances from medicinal plants. *World J. Diabetes*, 7(20): 605.
- Oz, AT and Kafkas E (2017). Phytochemicals in druits and vegetables. Superfood and functional food An overview of their processing and utilization, InTech.
- Padayatty SJ, Katz A, Wang Y, Eck P, Kwon O, Lee JH, Chen S, Corpe C, Dutta A and Dutta SK (2003). Vitamin C as an antioxidant: Evaluation of its role in disease prevention. J. Am. Coll. Nutr., 22(1): 18-35.
- Peters E, Anderson R, Nieman D, Fickl H and Jogessar V (2001). Vitamin C supplementation attenuates the increases in circulating cortisol, adrenaline and anti-inflammatory polypeptides following ultramarathon running. *Int. J. Sports Med.*, **22**(07): 537-543.
- Pocasap P, Weerapreeyakul N and Barusrux S (2013). "Cancer preventive effect of Thai rat-tailed radish (*Raphanus sativus* L. var. caudatus Alef). *J. Funct. Foods*, **5**(3): 1372-1381.
- Prahoveanu E and Eşanu V (1986). Immunomodulation with natural products. I. Effect of an aqueous extract of *Raphanus sativus* niger on experimental influenza infection in mice. *Virologie*, **38**(2): 115-120.
- Rao SS and R Najam (2016). Coconut water of different maturity stages ameliorates inflammatory processes in model of inflammation. *J. Intercult. Ethnopharmacol.*, **5**(3): 244-249.

- Roginsky V and Lissi EA (2005). Review of methods to determine chain-breaking antioxidant activity in food. *Food Chem.*, **92**(2): 235-254.
- Salah-Abbès JB, Abbès S, Ouanes Z, Houas Z, Abdel-Wahhab MA, Bacha H and Oueslati R (2008). Tunisian radish extract (*Raphanus sativus*) enhances the antioxidant status and protects against oxidative stress induced by zearalenone in Balb/c mice. *J. Appl. Toxicol.*, **28**(1): 6-14.
- Sham TT, Yuen ACY, Ng YF, Chan CO, Mok DKW and Chan SW (2013). A review of the phytochemistry and pharmacological activities of raphani semen. *eCAM*, **1**(1): 1-16.
- Siddiq A and Younus I (2018). The Radish, *Raphanus sativus* L. Var. caudatus reduces anxiety-like behavior in mice. *Metab. Brain Dis.*, **33**(4): 1255-1260.
- Slavin JL and Lloyd B (2012). Health benefits of fruits and vegetables. *Adv Nutr.*, **3**(4): 506-516.
- Songsak T and Lockwood G (2002). Glucosinolates of seven medicinal plants from Thailand. *Fitoterapia.*, **73**(3): 209-216.
- Takaya Y, Kondo Y, Furukawa T and Niwa M (2003). Antioxidant constituents of radish sprout (kaiwaredaikon), *Raphanus sativus* L. *J. Agric. Food Chem.*, **51**(27): 8061-8066.
- Thejass P and Kuttan G (2007). Immunomodulatory activity of Sulforaphane, a naturally occurring isothiocyanate from broccoli (*Brassica oleracea*). *Phytomedicine*, **14**(7): 538-545.
- Younus I and Siddiq A (2017). Behavioral evidence of antidepressant-like activity of *Raphanus sativus* L. var. caudatus in mice. *Afr. J. Tradit. Complement. Altern. Med.*, **14**(3): 142-146.
- Younus I and Siddiq A (2022). *Raphanus sativus* L. Var. caudatus as an analgesic and antipyretic agent in animal models. *Pak. J. Zool.*, **54**(4): 1643.